

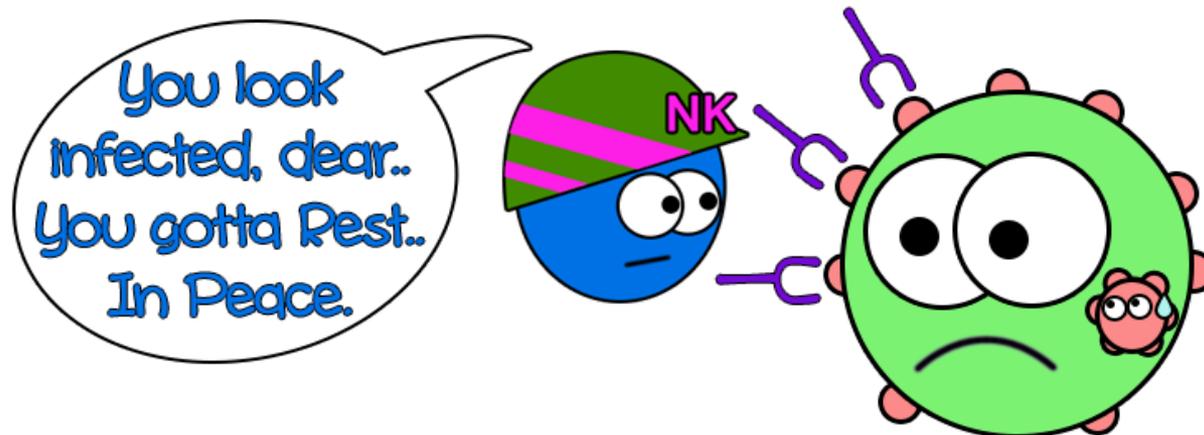


**INNO MOL**  
Innovation Pipeline

# High throughput screening of phage display libraries for production of fully human antibodies challenged to cells expressing native Claudin-1

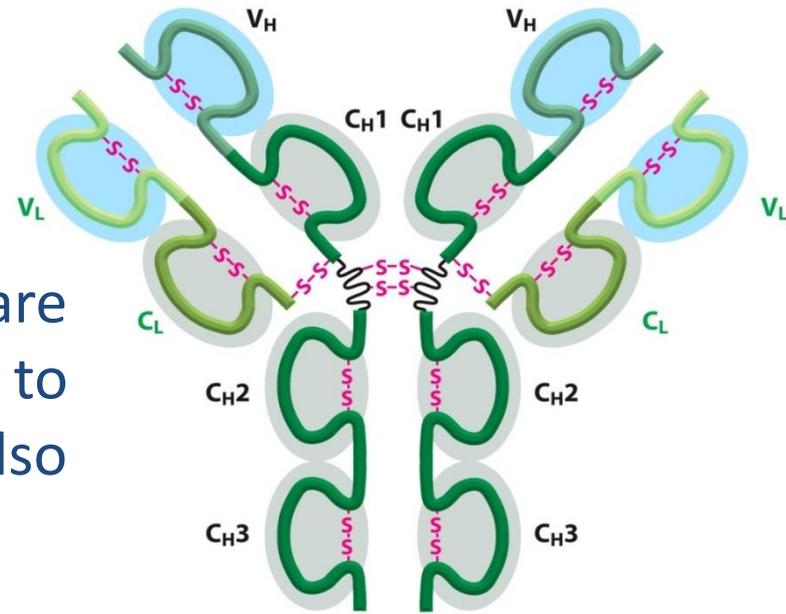
Emanuele Sasso

Genomics and Bioinformatics Workshop 11.2015



# The Structure of Antibodies

Immunoglobulins of class G (IgG) are constituted by two heavy chains H equal to each other, and two light chains L also equal to each other.

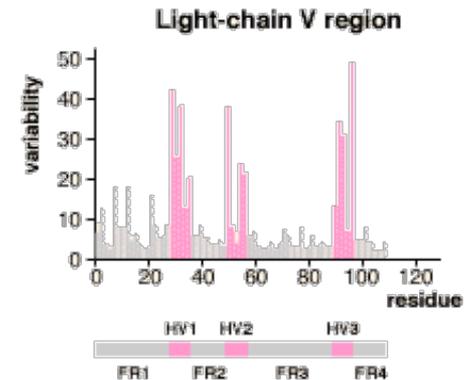
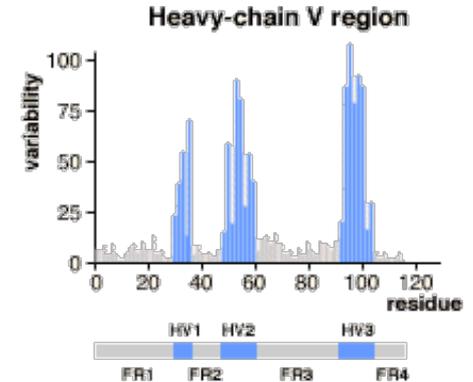
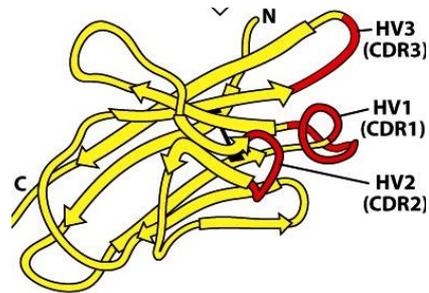
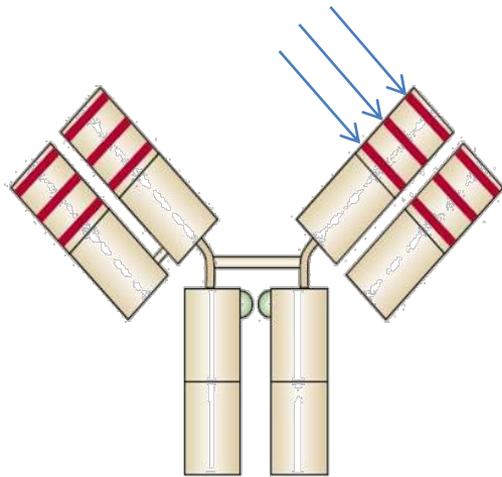


The L and H chains are composed of variable domains (VL and VH) and constant domains, one for the L chain (CL), and three for the H chain (CH1 CH2 CH3)

**The protein domains responsible for antigen recognition are in the variable regions VH and VL**

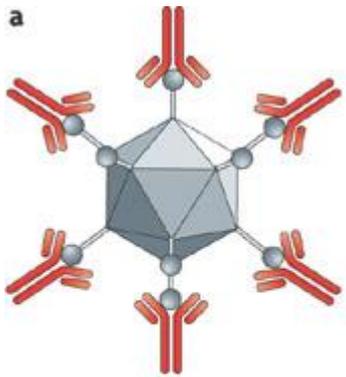
# The Structure of Antibodies

Hypervariable domains  
(complementarity determining regions CDR)

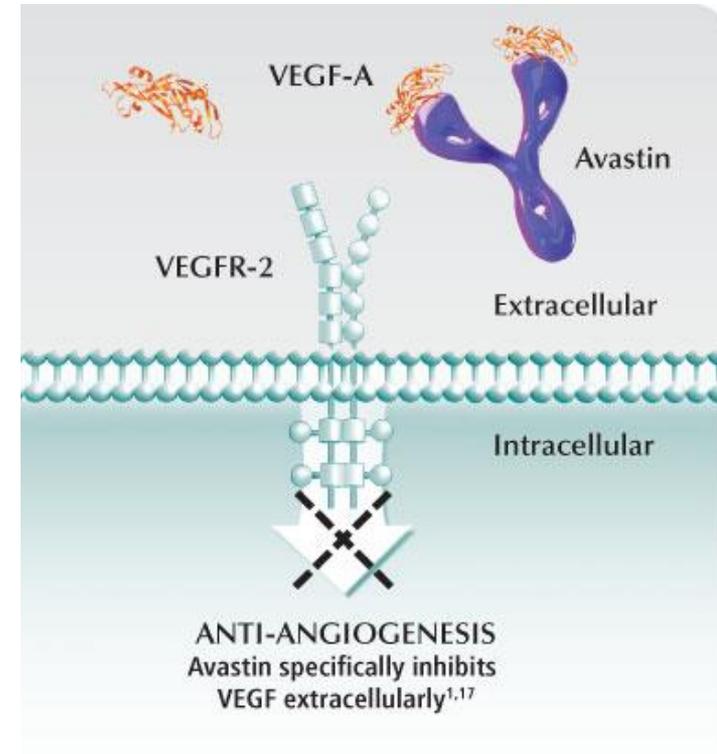
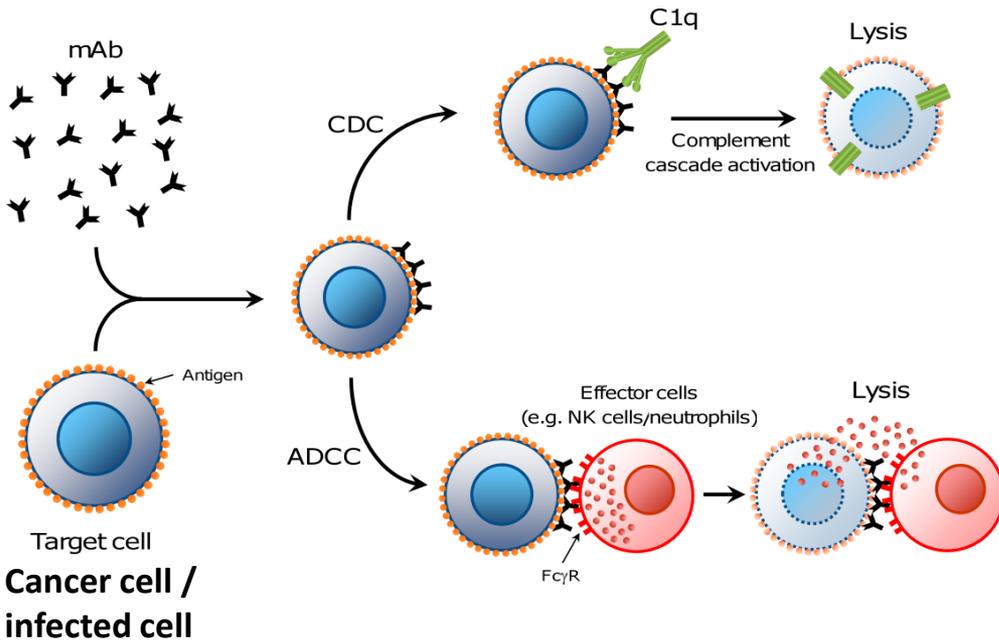
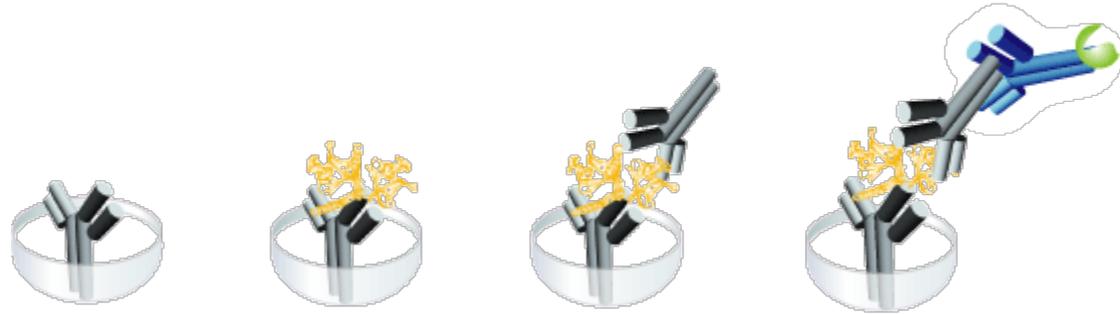


The variable region of an antibody is responsible of its binding to an epitope. The variable region is further subdivided into hypervariable (HV) and framework (FR) regions. Within light and heavy chains, exist three hypervariable regions and four FR regions. The hypervariable regions are definitely involved in epitope binding.

# Application of recombinant monoclonal antibodies

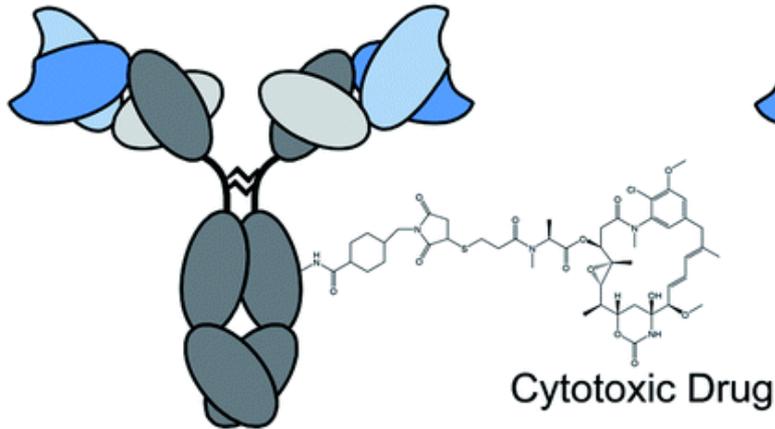


Virus neutralized by antibodies

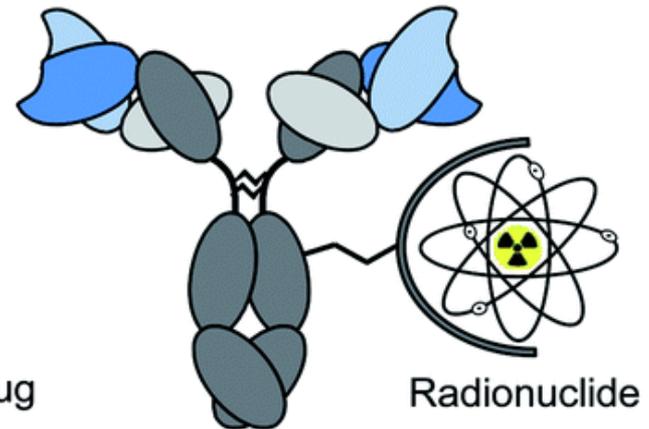


# Advanced applications

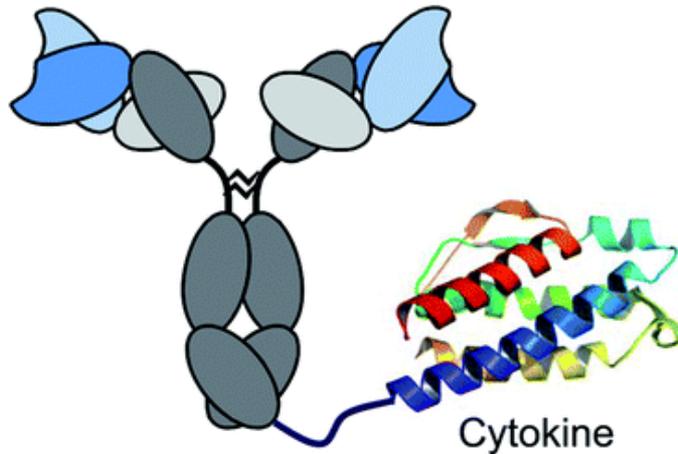
## Antibody-Drug Conjugate



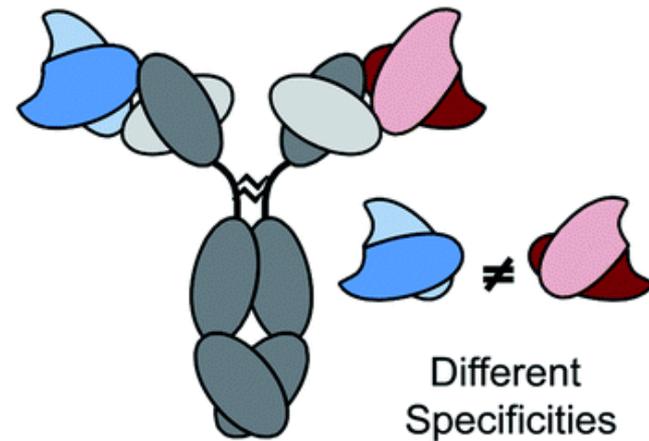
## Radioimmunoconjugate



## Immunocytokine

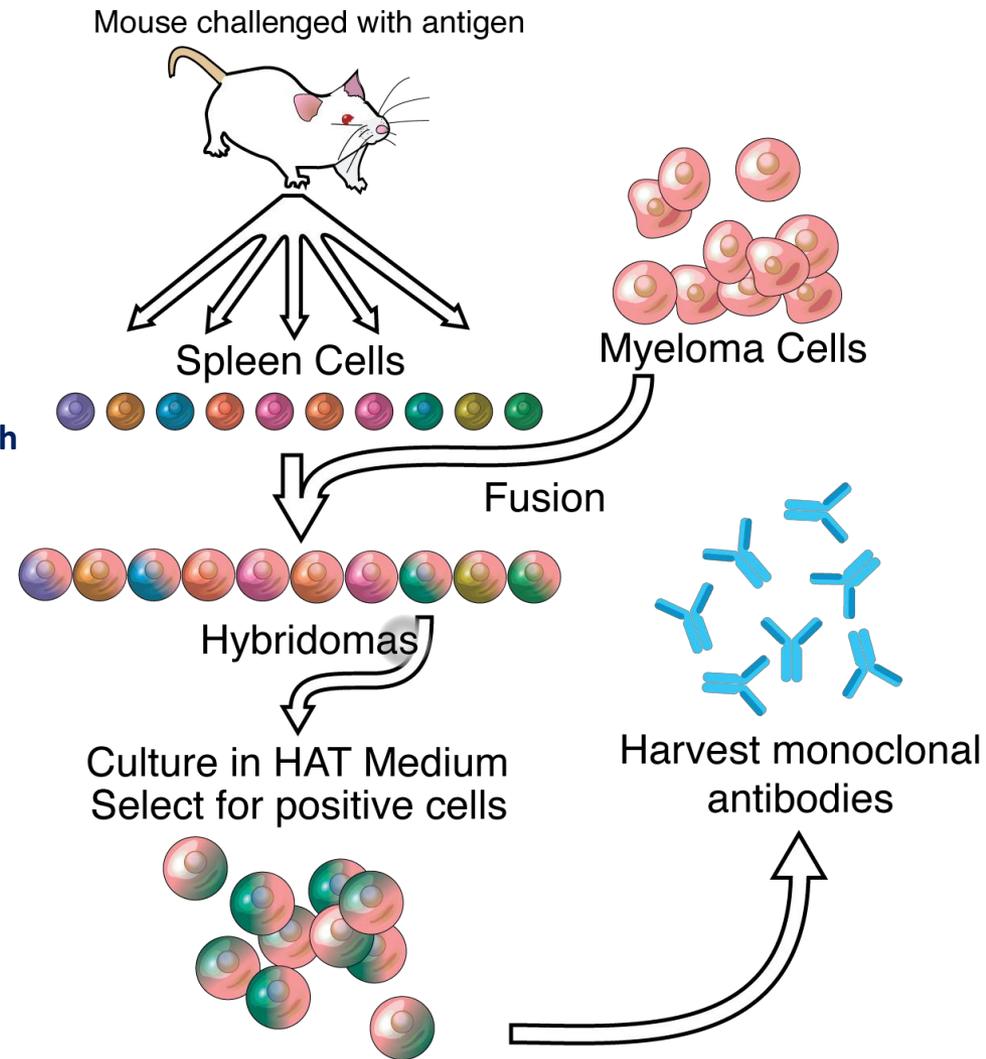


## Bispecific Antibody



# Murine antibodies by hybridoma technique

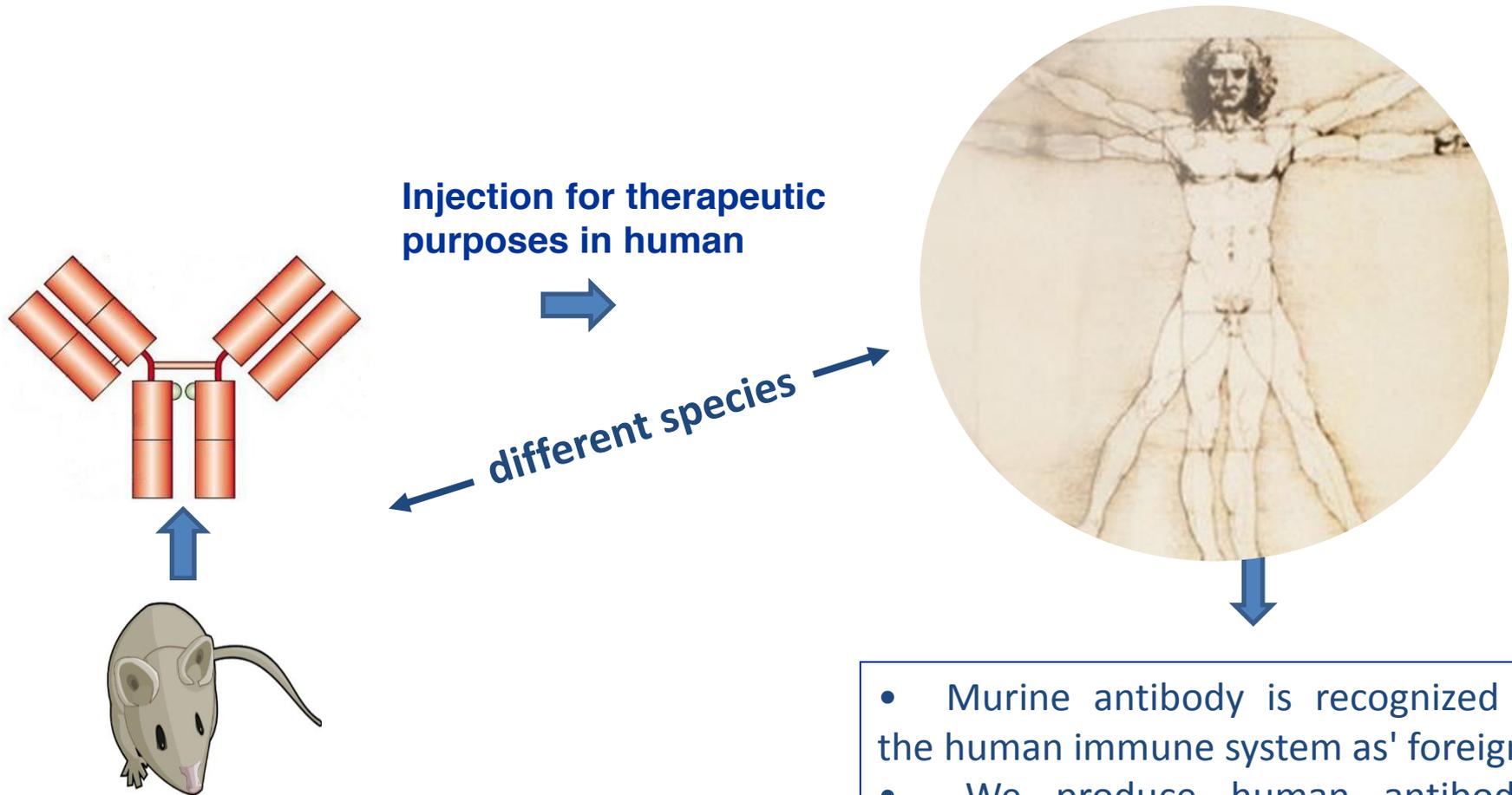
- 1) Immunize animal (mouse or rabbit)
- 2) Isolate spleen cells (containing antibody-producing B cells)
- 3) Fuse spleen cells with myeloma cells (e.g. using PEG - polyethylene glycol)
- 4) Allow unfused B cells to die
- 5) Add HAT culture to kill unfused myeloma cells
- 6) Clone remaining cells (place 1 cell per well and allow each cell to grow into a clone of cells)
- 7) Screen supernatant of each clone for presence of the desired antibody (ELISA)



# Problems with using mouse mAb

- The therapeutic use of rodent monoclonal antibodies in humans is limited by their immunogenic, short circulating half-life, and inability to efficiently trigger human effectors mechanisms:
  - This is due to differences between the mouse and humans.
  - Also severe allergic response in human when mouse mAb are introduced to a patients.
  - Also constant region of murine mAb are not effective in interacting with human effectors molecules.

# Induction of human anti-murine mAb : HAMA (Human Anti-Mouse Antibodies)

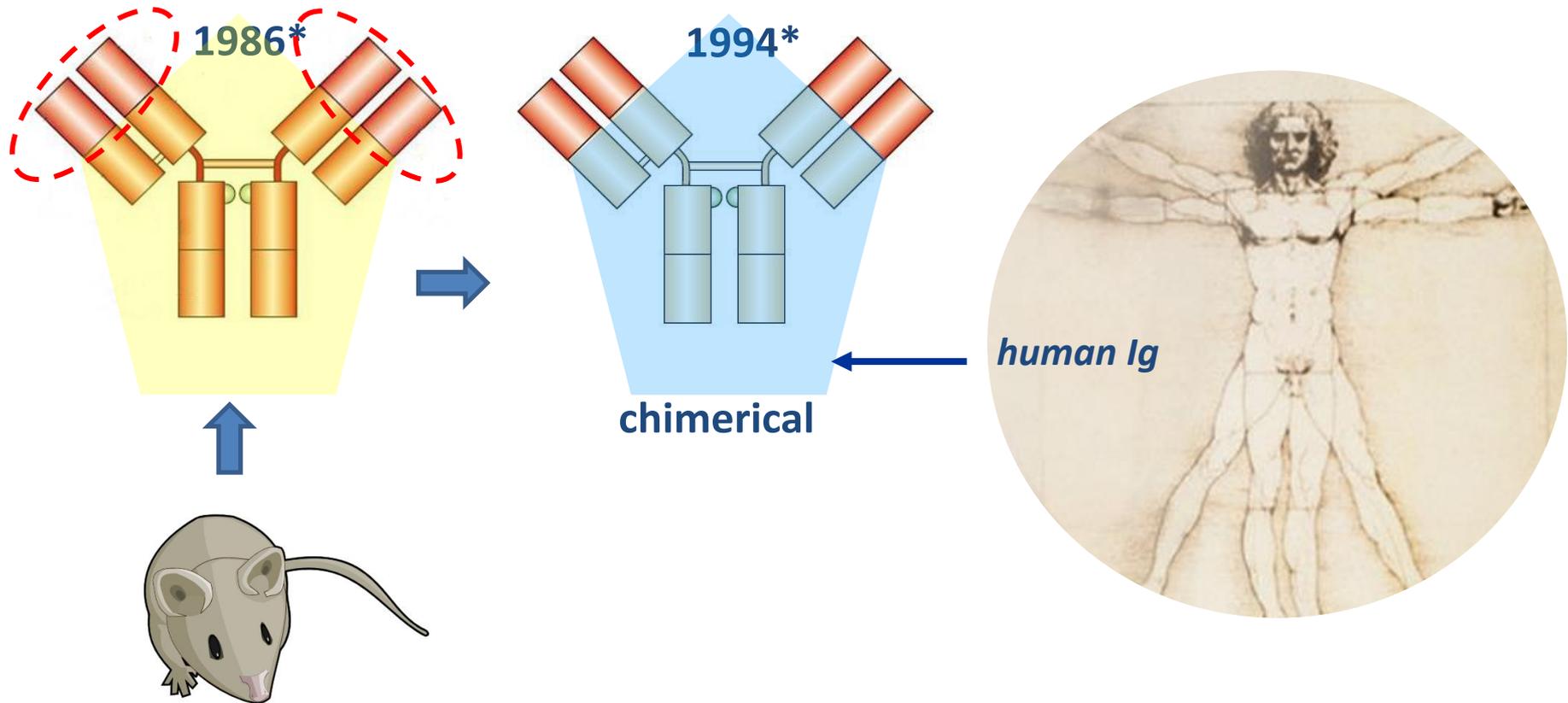


- Murine antibody is recognized by the human immune system as 'foreign'
- We produce human antibodies against murine mAb that is inactivated
- Therapy becomes ineffective

# Generation of **chimeric** monoclonal antibodies

The solution:

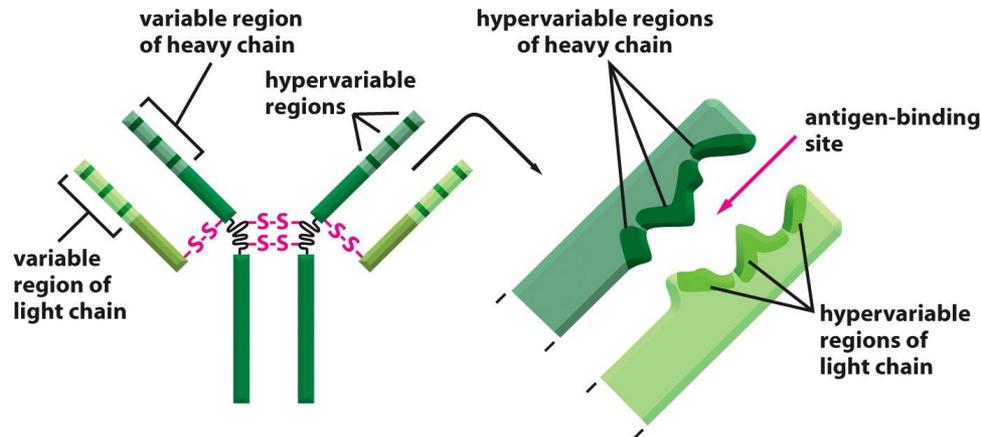
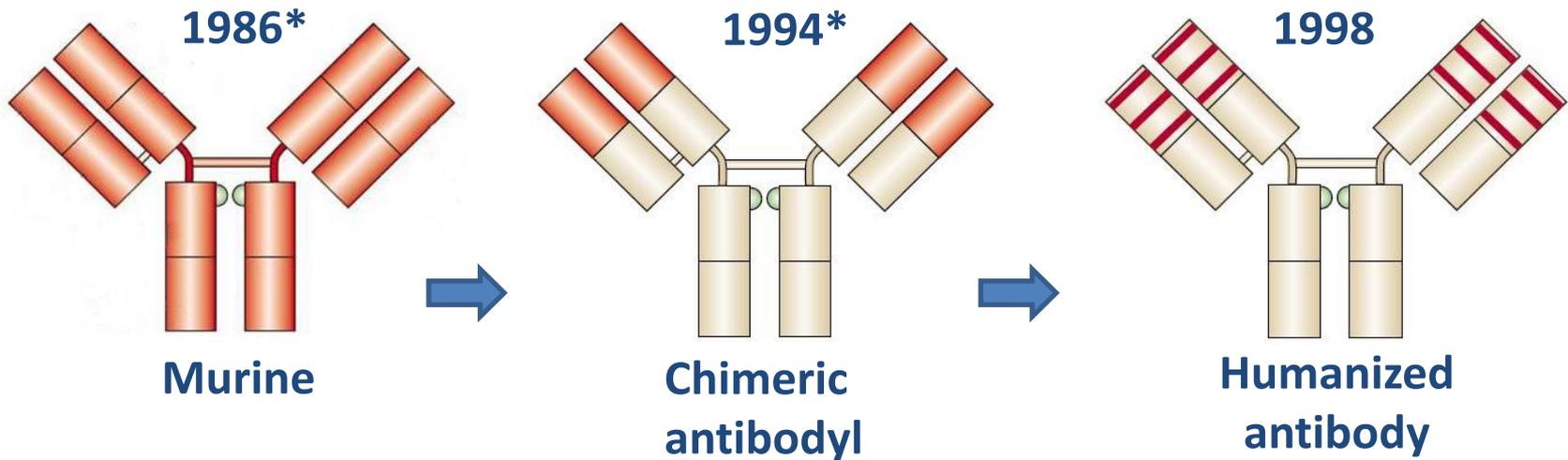
- Replace the constant part (CH) of the mouse antibody with that of a human one.



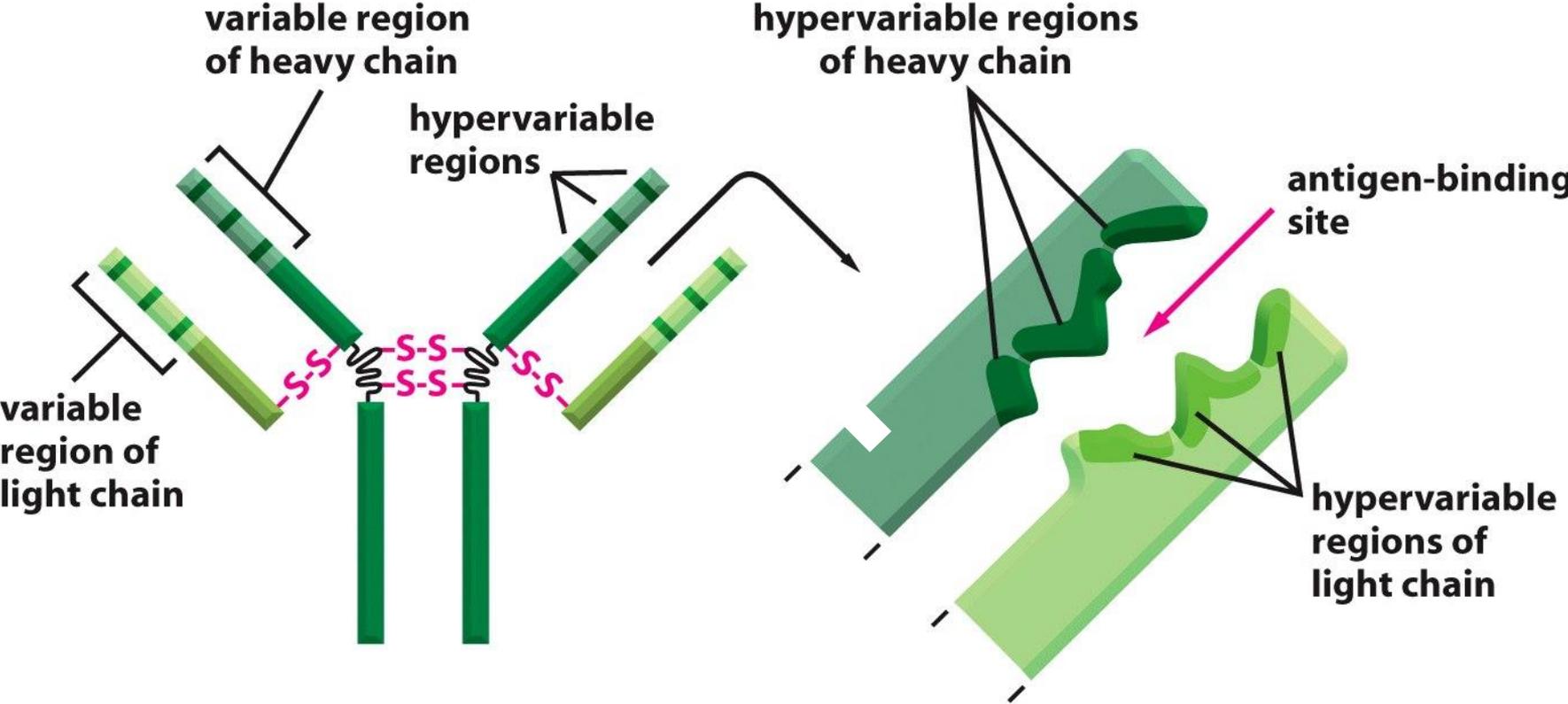
# From chimeric to **humanized** antibody

The solution:

- 'Transplant' only the hypervariable murine antibody in a human antibody  
= Humanized antibody

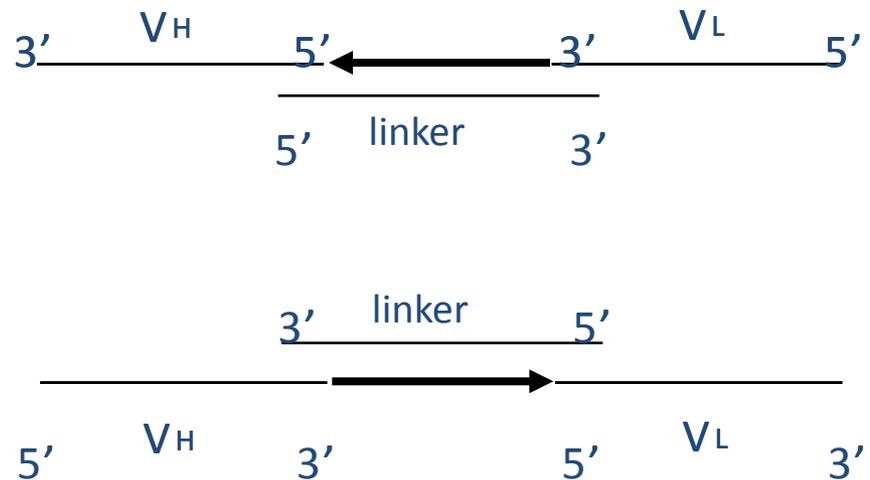
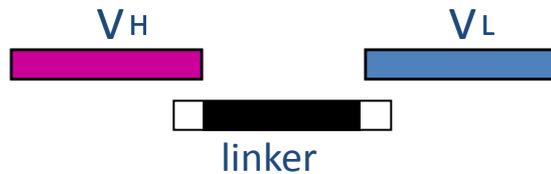


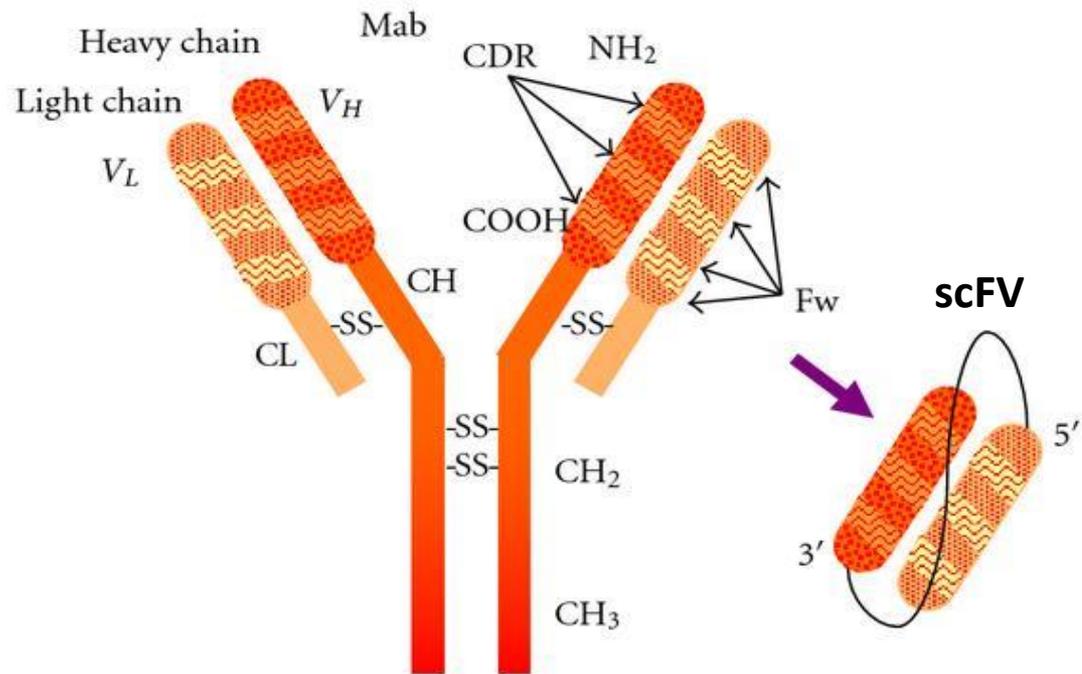
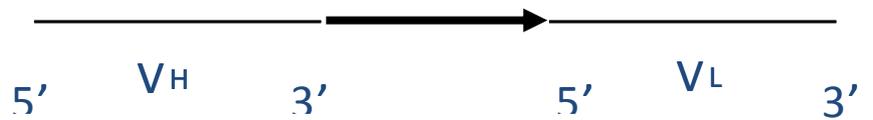
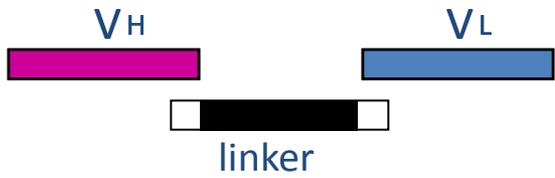
# The only variable regions are needed for epitope binding



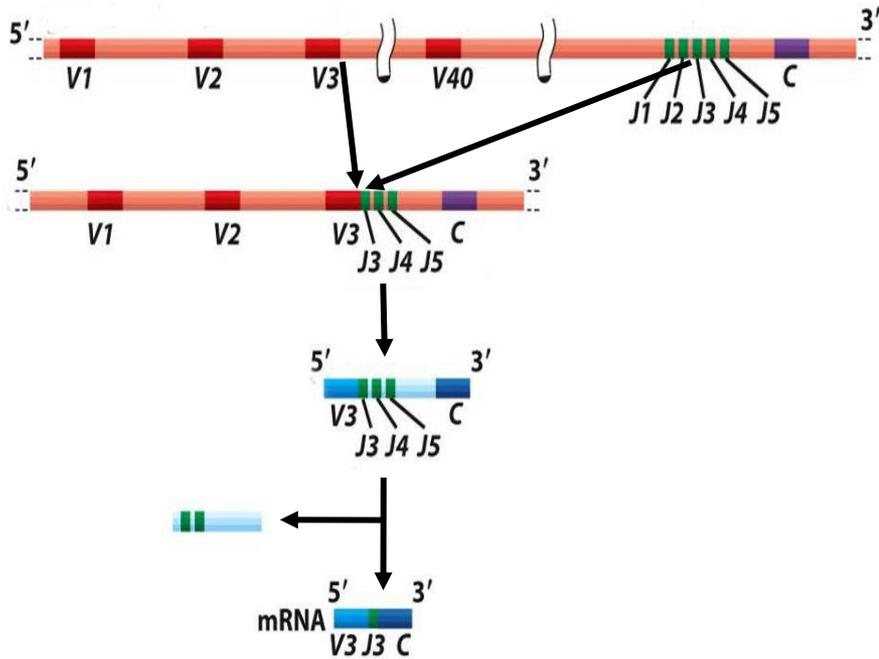
# Random combination of $V_H$ - $V_L$ by flexible linker

- $V_H$  and  $V_L$  are amplified and combined through a polypeptidic linker (Gly 4, Ser 3)





# In vivo vs in vitro diversity



**$10^8$**  in vivo immune  
system different  
antibodies

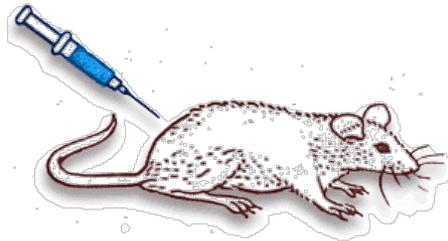


**$10^9-10^{11}$**  ScFv

# Where from to isolate VH and VL?

## Immunised library:

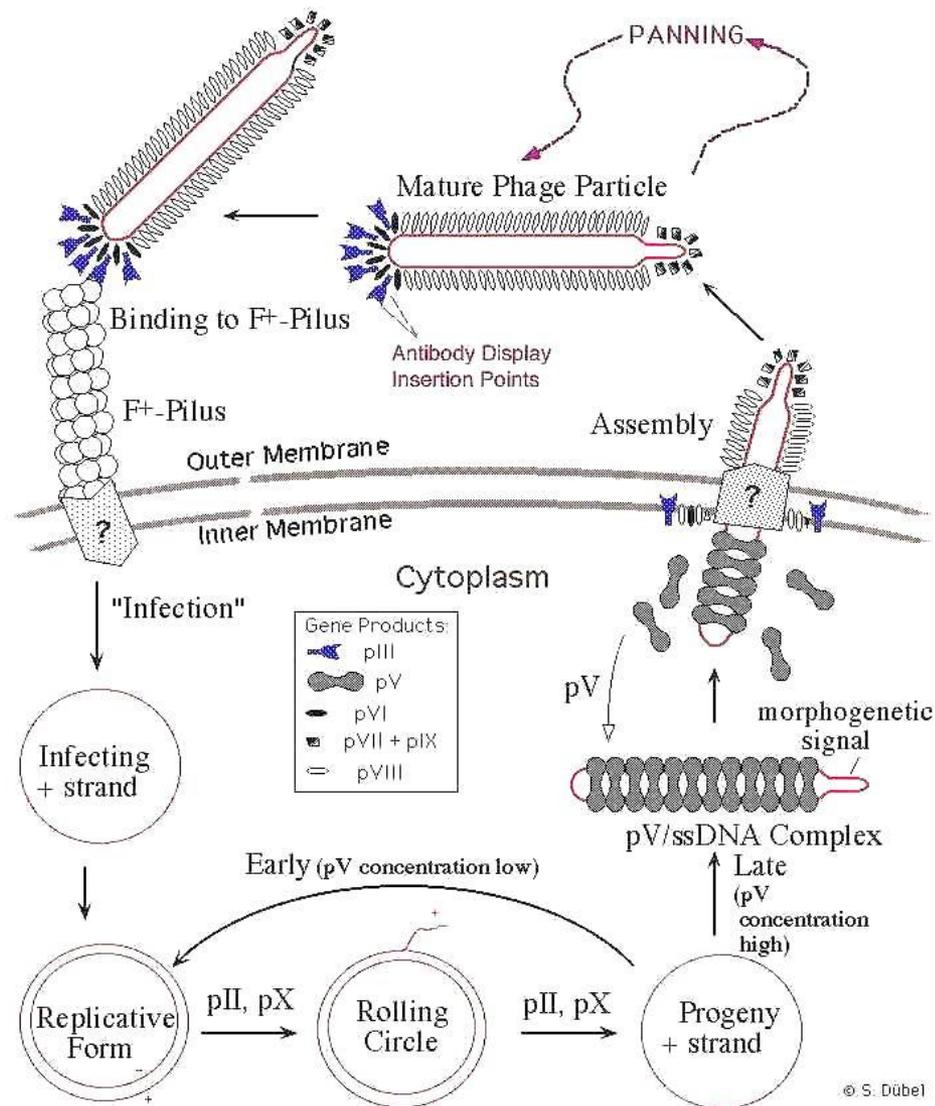
- VH and VL are isolated from B cells of an immunised animal or humans.
- Suitable for difficult antigens



## Naive library/germline library:

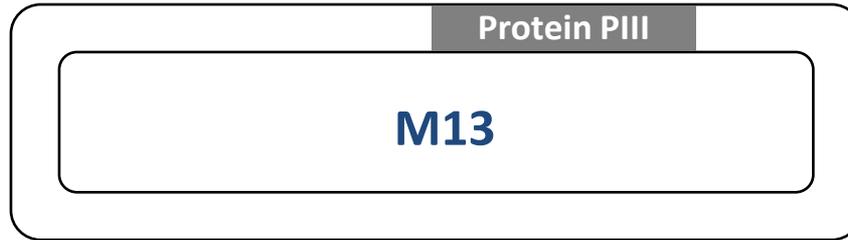
- VH and VL are isolated from healthy **human** donors and randomly combined.
- Suitable for almost all antigens

# Life cycle of filamentous phage

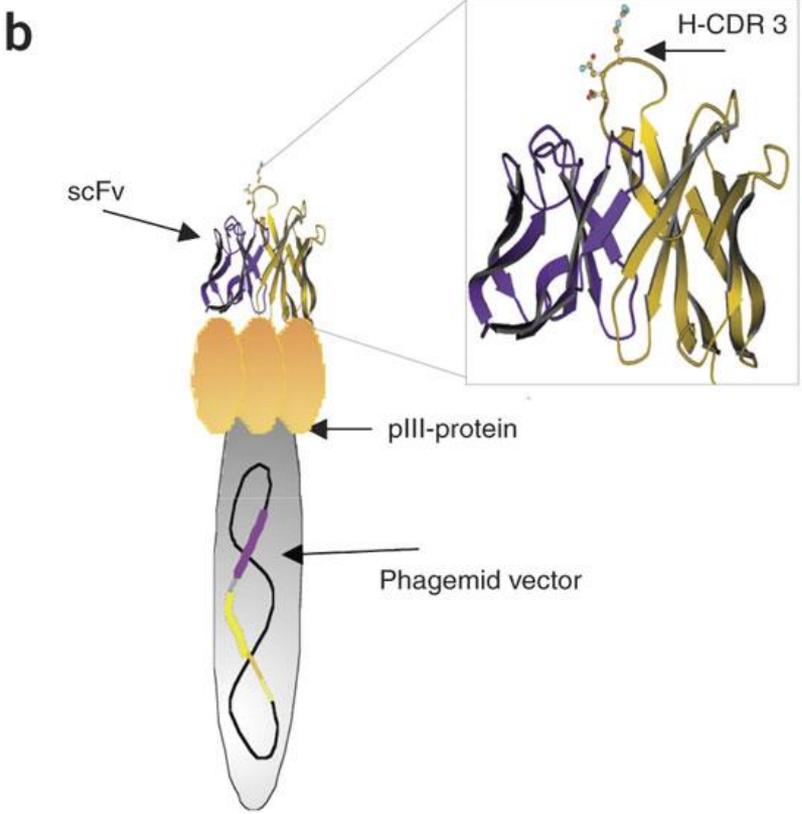
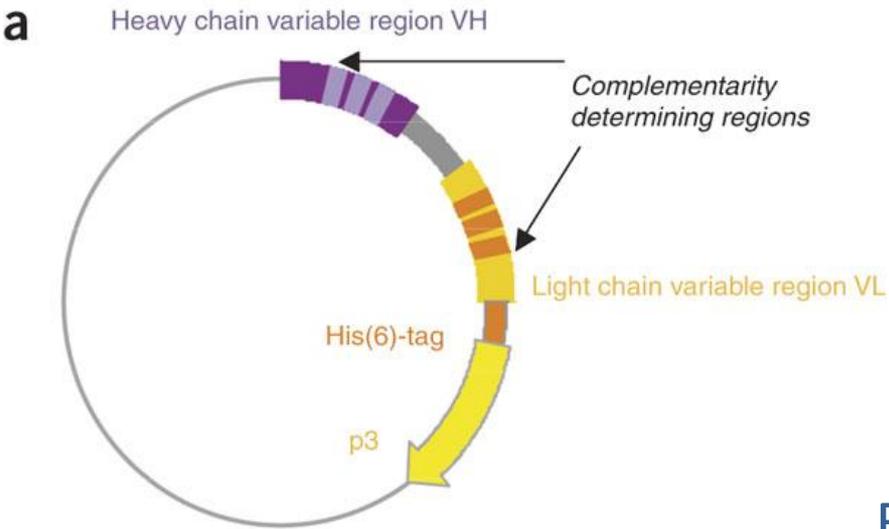


# How to screen the scfvs?

## Phage Display



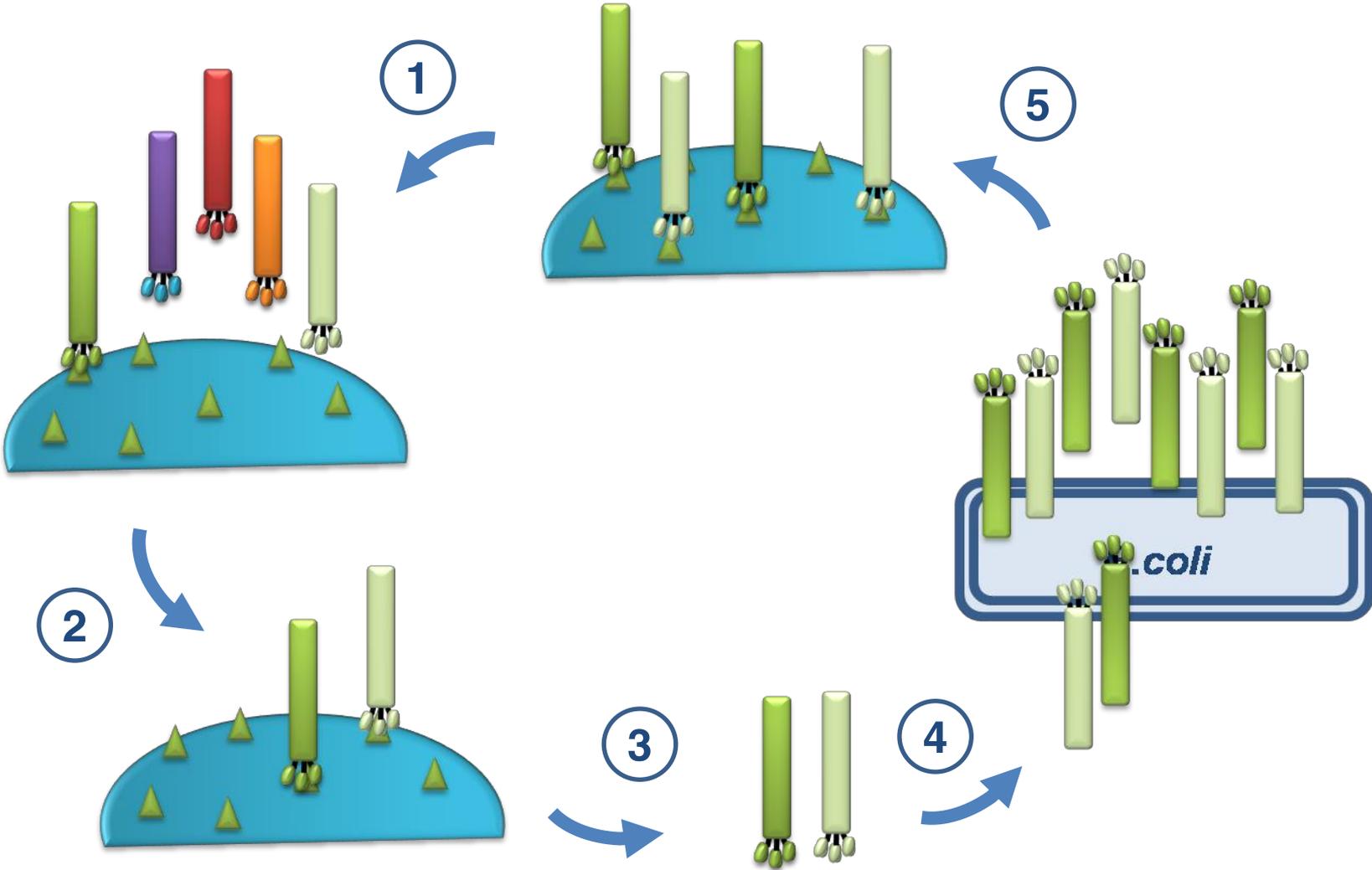
Scfv are “exposed” on M13’s capsid in frame fused to PIII protein



Each phage bring together the scfv protein and its «genetical information»

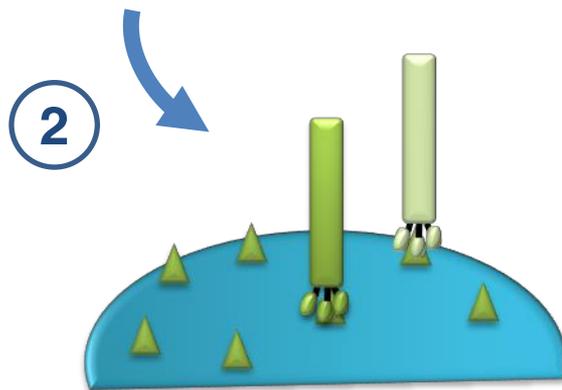
**1phage = 1ScFv**

# Phage display





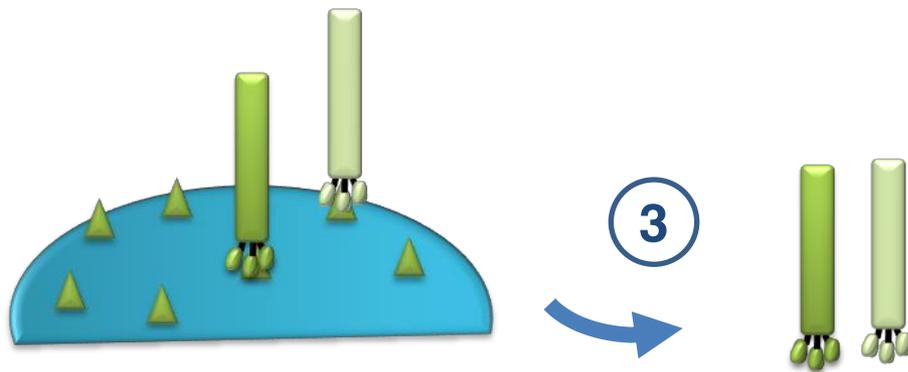
1. The scfv/phage library is incubated with epitope



2.

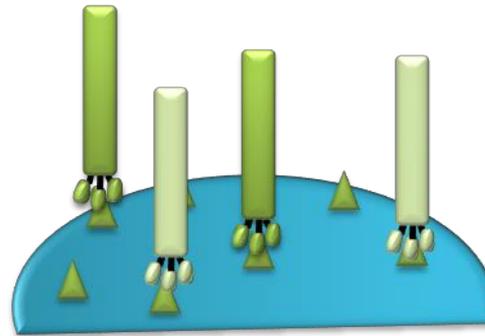
-The non-specific ScFv/phages are washed up.

-Specific ScFv/phage stay on epitope



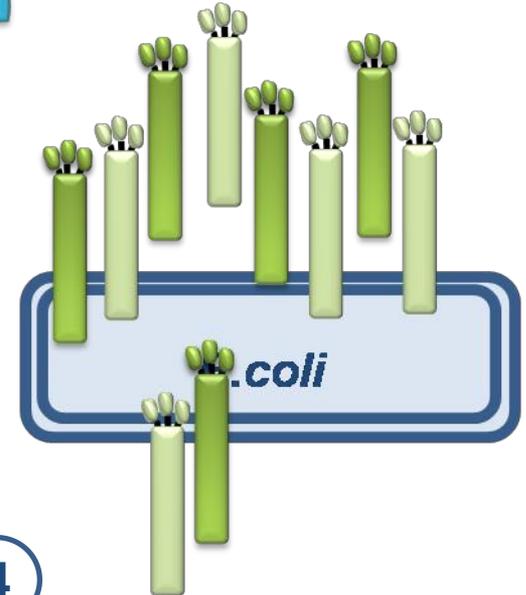
3. The phage bound are eluted (pH, trypsin)

5. Amplified phages are incubated with epitope

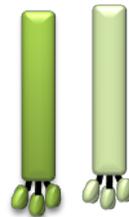


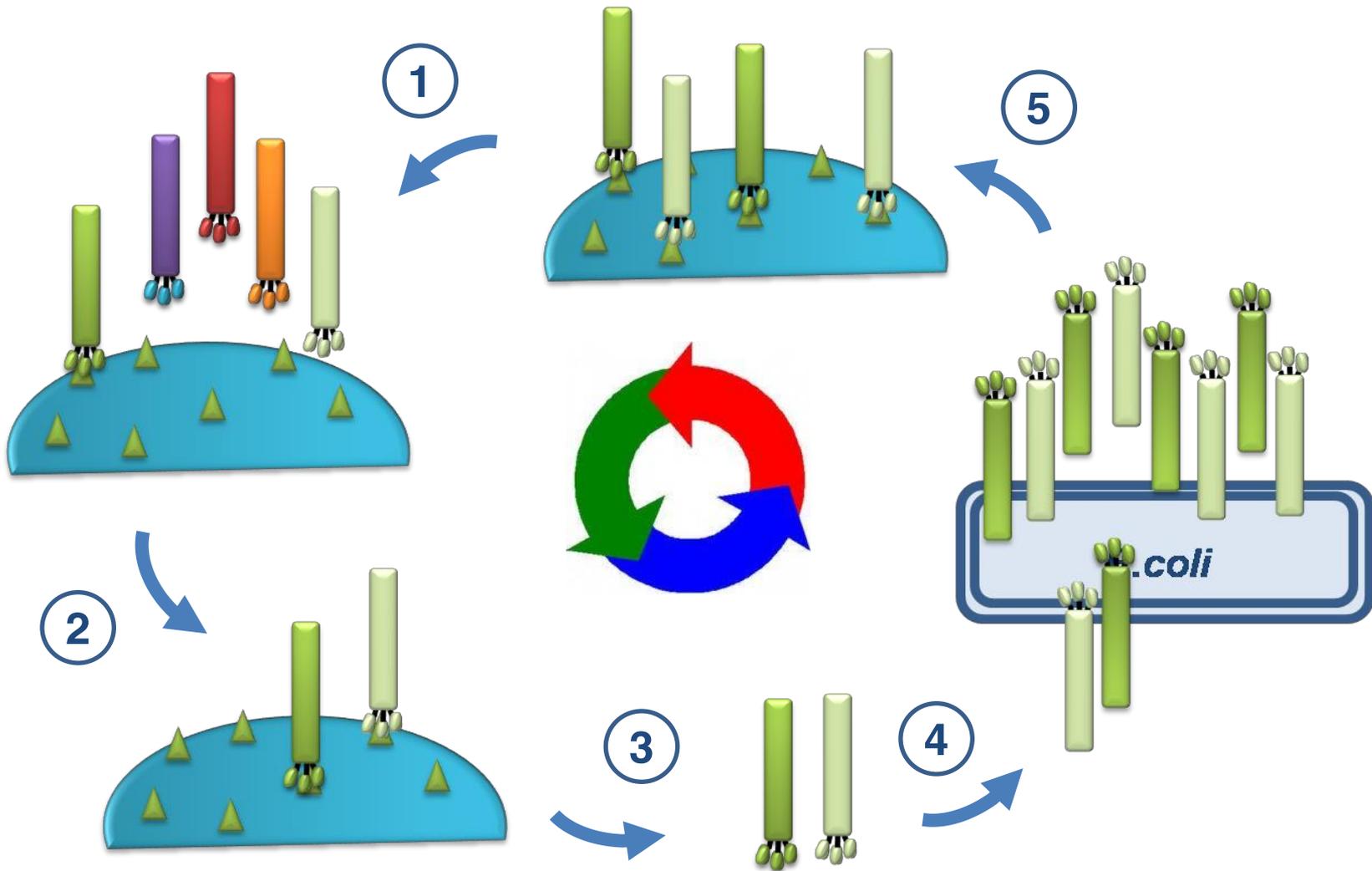
5

4. E. Coli are infected with eluted phages in order to amplify the selected ScFvs

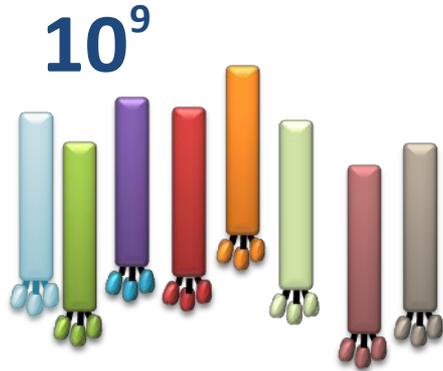


4

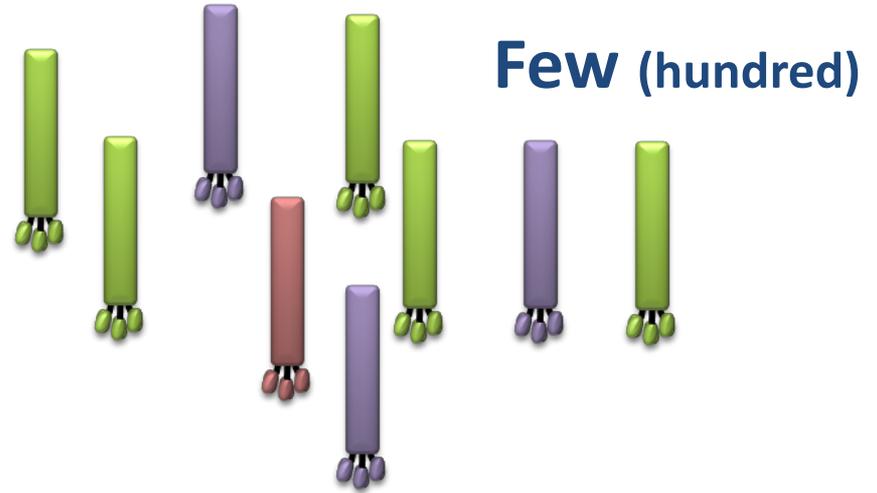




# From protein to DNA

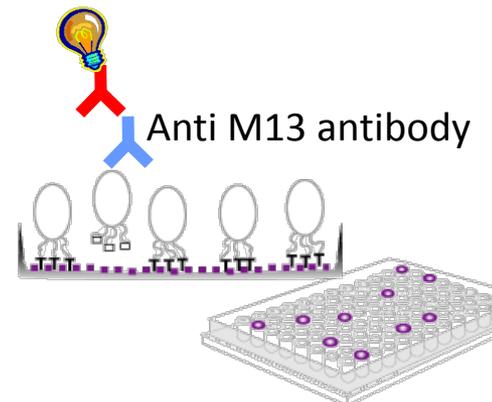


Selection  
cycles  
→



Random phages are  
tested by ELISA

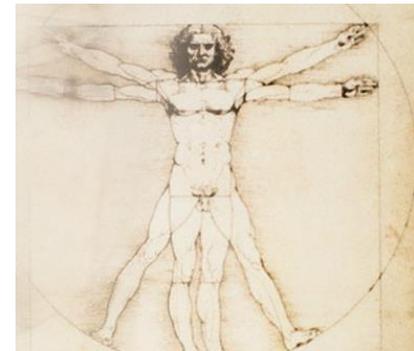
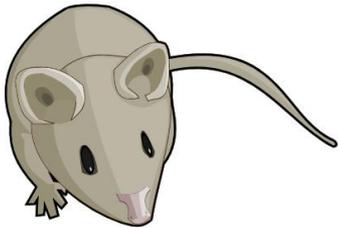
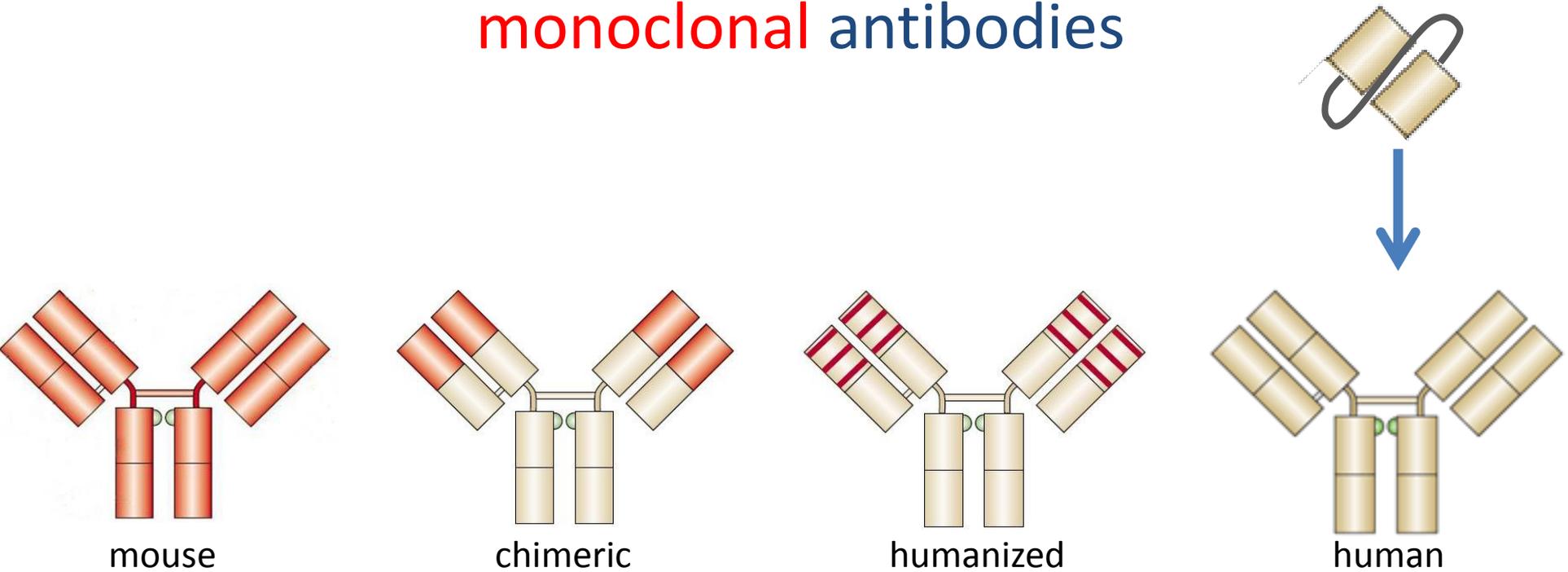
Positive phages  
from ELISA are  
sequenced



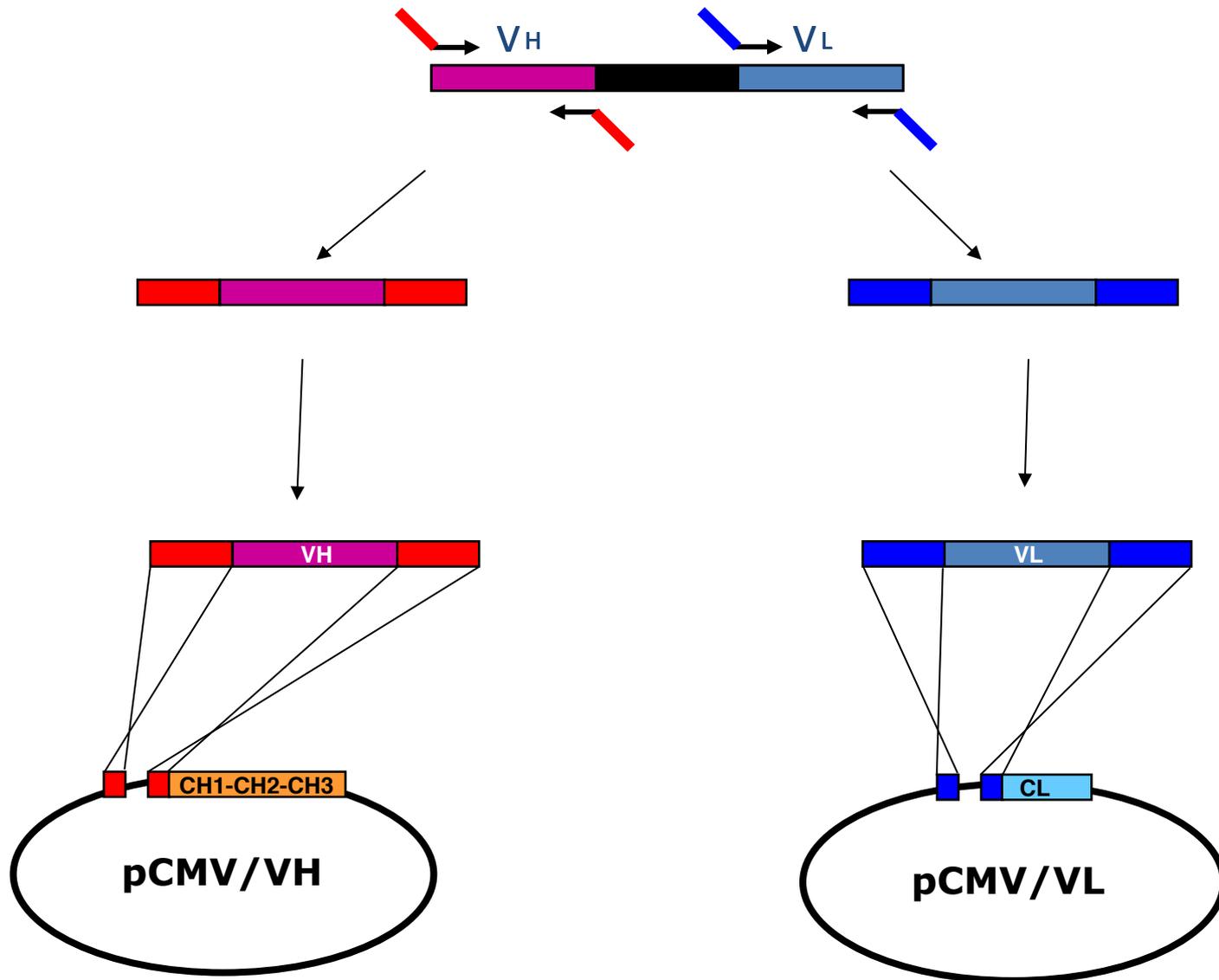
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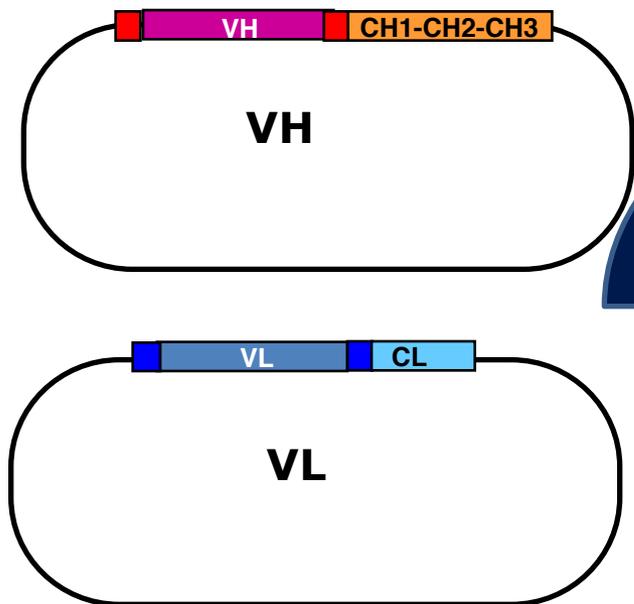
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CTGGGGGGTCCCTGAGACTCTCCTGTGCAGCCTCTGGATTACCTTTAGCAGCTATGCCATGAGCTGGGTCCGCCAG
GCTCCAGGGAAGGGGCTGGAGTGGGTCTCAGCTATTAGTGGTAGTGGTGGTAGCACATACTACGCAGACTCCGTGAAG
GGCCGGTTCACCATCTCCGGAGACAATCCAAGAACACGCTGTATCTGCAAAATGAACAGCCTGAGAGCCGAGGACACG
GCCGTGTATTACTGTCAAGGTCGAACTGGTACTTCGATCTCTGGGGCAAAGGGACAATGGTCACCGTCTCGAGTGG
GGCGCGGTTCAGCGGAGGTGGCTCTGGCGGTGGCGGAAGTGCACITTCCTCTGAGCTGACTCAGGACCCTGCTGTG
TCTGTTGCCCTTGGGACAGACAGTCAAGCTCACATGCCGAGGAGACAACCTCAGAAGTTATTATGCCAGCTGGTACCAG
CAGCGGCCAGGACAGGCCCTAGACTTGTCAATGATGGGAAAAACATCCGGCCCTCAGGGATCCCAGACCGATTCTCT
GGCTCCAGCTCAGGGACACAGGTTTCGTTGACCATCACTGGGGCTCAGCGGAGGATGAGGCTGACTATTACTGCAAC
TCCCGGGACAGCCTTACTAACCGTCCCTTTTCGGCGGAGGGACCAAGGTCACCGTCTTAGGTGCGGCCGCACATCAT
CATCACCATCAGGGGCCGAGAACAAAACACTCATCTCAGAGAGGATCTGAATGGGCCCC
    
```

# ScFvs allow to produce fully **human** **monoclonal** antibodies

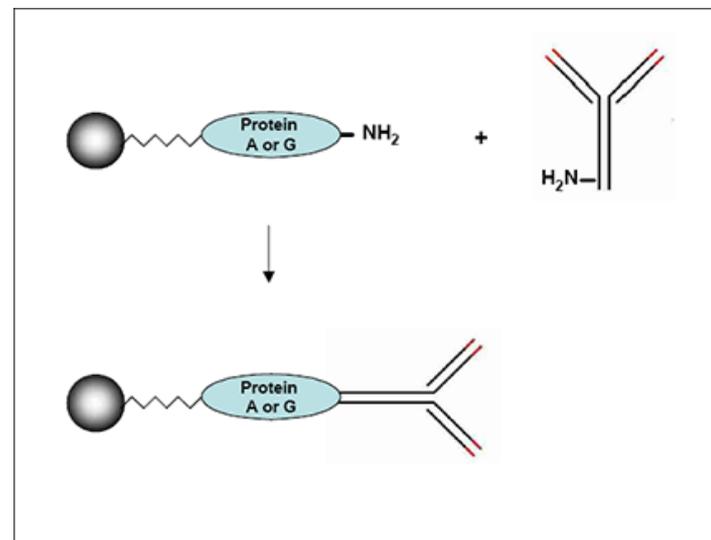
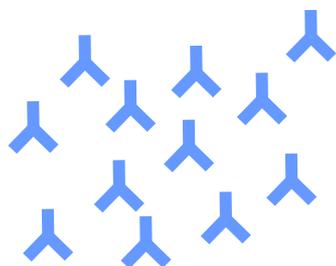
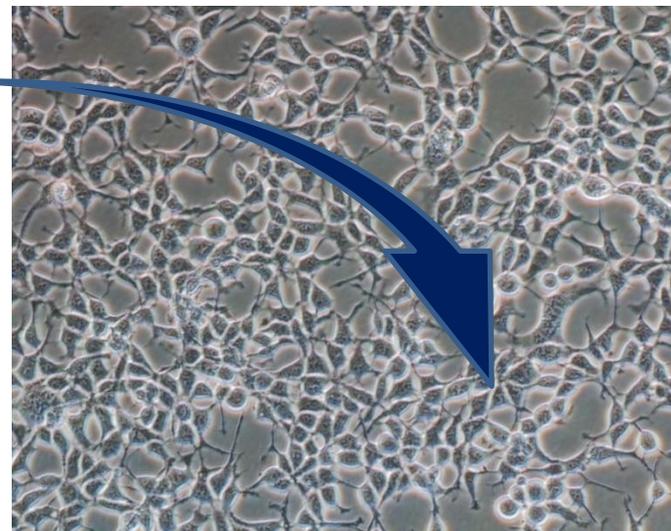


# From ScFv to human antibody



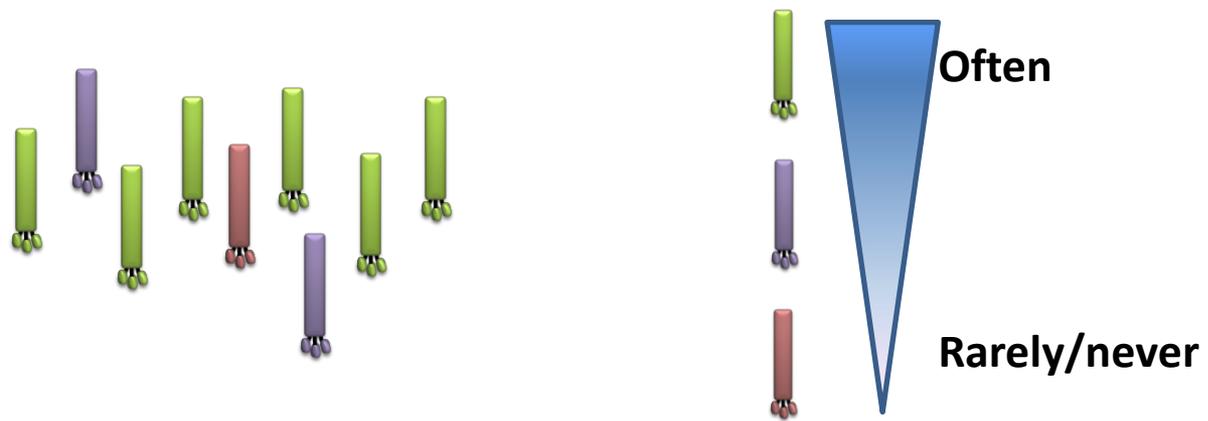


Co-transfection



# Problems related to classical ELISA screening of phages libraries:

- Repetitive isolation of the same clones within and between selection steps



- Information in phage clones enrichment needs redundant sequencing after each step

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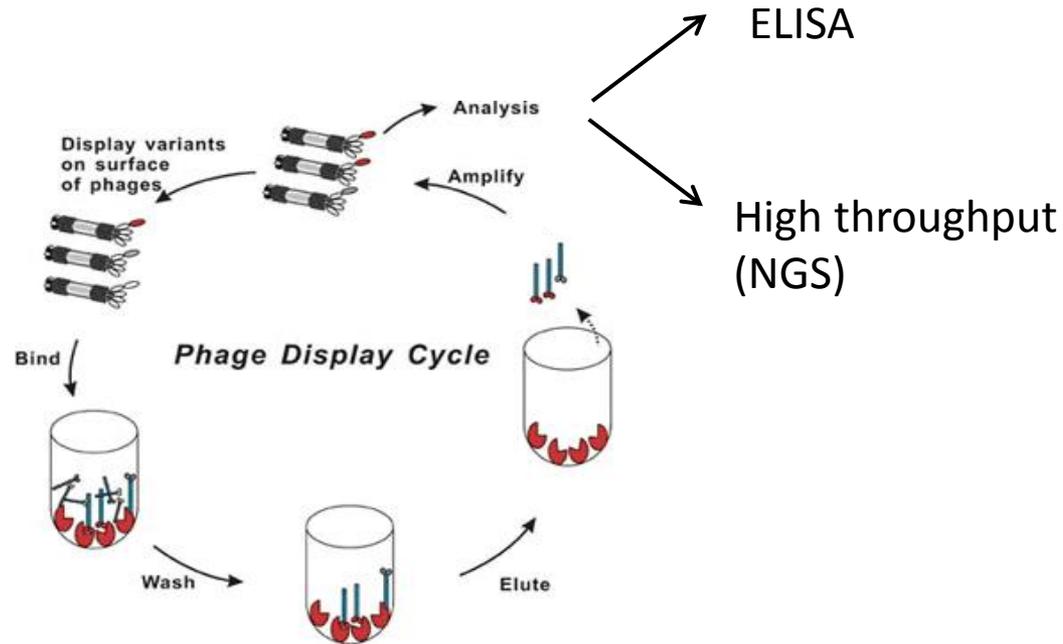
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5      CGACCTGGAAAGCGGGCAGTGGAGCGCAACGCAATTAATGTTGAGTTTAGCTCACTCATT
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7      CGACCTGGAAATATAGGCAGTGGAGCGGGACGCAATTAATGTCACGTTTAGCTCACTCATGT
8      CGACCTGGAAATATAGGCAGTGGAGCGGGACGCAATTAATGTCACGTTTAGCTCACTCATGT
9      CGACCTGGAAAGCGGGCATACTCUGGCAACGCAATTATTCGGTGTTAGCTCACTCATA
*****          ****          *  *****  *          *****
    
```

- Continuous handling of clones during selection steps (isolation, growth, ELISA)

# Cost per Genome



# Selecting scFvs from phage libraries by next generation sequencing



The **deep sequencing approach** could **identify under-represented clones** in phage screens. It could also preserve diversity of phage clones and identify ligands previously lost in phage display screens.

# How do we sequence scfv?

ATGAAATACC	TATTGCCTAC	GGCAGCCGCT	GGATTGTTAT	TACTCGCGGC	CCAGCCGG	CC	60
ATGG	CCGAGG	TGCAGCTGGT	GGAGTCTGGG	GGAGGCGTGG	TCCAGCCTGG	GAGGTCCCTG	120
AGACTCTCCT	GTGCAGCCTC	TGGATTCACC	TTCAGTAGCT	ATGCTATGCA	CTGGGTCCGC		180
CAGGCTCCAG	GCAAGGGGCT	GGAGTGGGTG	GCAGTTATAT	CATATGATGG	AAGCAATAAA		240
TACTACGCAG	ACTCCGTGAA	GGGCCGATTC	ACCATCTCCA	GAGACAATTC	CAAGAACACG		300
CTGTATCTGC	AAATGAACAG	CCTGAGAGCT	GAGGACACGG	CCGTGTATTA	CTGTGCAAGA		360
GGGTTTCCTA	TGCCGTGGGG	CCAAGGTACC	CTGGTCACCG	TGTCGAGAGG	TGGAGGCGGT		420
TCAGGCGGAG	GTGGCTCTGG	CGGTGGCGGA	TCGTCTGAGC	TGACTCAGGA	CCCTGCTGTG		480
TCTGTGGCCT	TGGGACAGAC	AGTCAGGATC	ACATGCCAAG	GAGACAGCCT	CAGAAGCTAT		540
TATGCAAGCT	GGTACCAGCA	GAAGCCAGGA	CAGGCCCTG	TACTTGTCAT	CTATGGTAAA		600
AACAACCGGC	CCTCAGGGAT	CCCAGACCGA	TTCTCTGGCT	CCAGCTCAGG	AAACACAGCT		660
TCCTTGACCA	TCACTGGGGC	TCAGGCGGAA	GATGAGGCTG	ACTATTACTG	TAACTCCCGG		720
GACAGCAGTG	GTAACCATGT	GGTATTCGGC	GGAGGGACCA	AGCTGACCGT	CCTAGGT	GCG	780
GCCGCAGAAC	AAAAACTCAT	CTCAGAAGAG	GATCTGAAT				

-The whole scfv consists of about 800 bp. It is too long for NGS sequencing

-The separated sequencing of VH and VL would be too expensive and doesn't allow to associate the VH with the correspondent VL

# Which technology?



HiSeq 2500/2000



HiSeq 2500



MiSeq



Roche 454

Mode	High Output	Rapid Run		
Output (maximum)	600Gb	180Gb	8.5Gb	1 Gb
Run Time	2 - 11 Days	7 - 40 Hr.	4 - 39 Hr.	10-20Hr.
Paired-end Reads (maximum)	6 Billion $6 \times 10^{12}$	1.2 Billion $10^{12}$	15 Million $15 \times 10^6$	1 million $10^6$
Single-end Read (maximum)	3 Billion	600 Million	7.5 Million	
Max Read Length	2x100 bp	2x150 bp	2x250 bp	700 bp

MiSeq platform(Illumina) allows to cover the whole Heavy Variable region with a coverage at least of 100x

Advantages	Disadvantages
<ul style="list-style-type: none"> <li>•Cheap</li> <li>•Good coverage <b>10<sup>7</sup></b></li> </ul>	<ul style="list-style-type: none"> <li>•NGS doesn't allow to sequence the whole fragment of VH and VL</li> </ul>



MiSeq

Mode	
Output (maximum)	8.5Gb
Run Time	4 - 39 Hr.
Paired-end Reads (maximum)	15 Million
Single-end Read (maximum)	7.5 Million
Max Read Length	2x250 bp
Bases above Q30 (2x100bp)	>85% (2x100 bp)
Required Input	1ng with NexteraXT 50 ng with Nextera 100 ng – 1 µg with TruSeq

The VH is usually more involved than VL in epitope binding.

# Workflow of high throughput phage display selection

1. Perform 4 selection cycles by phage display
2. Excise VHs from the 4 sub-libraries of phages by restriction endonucleases

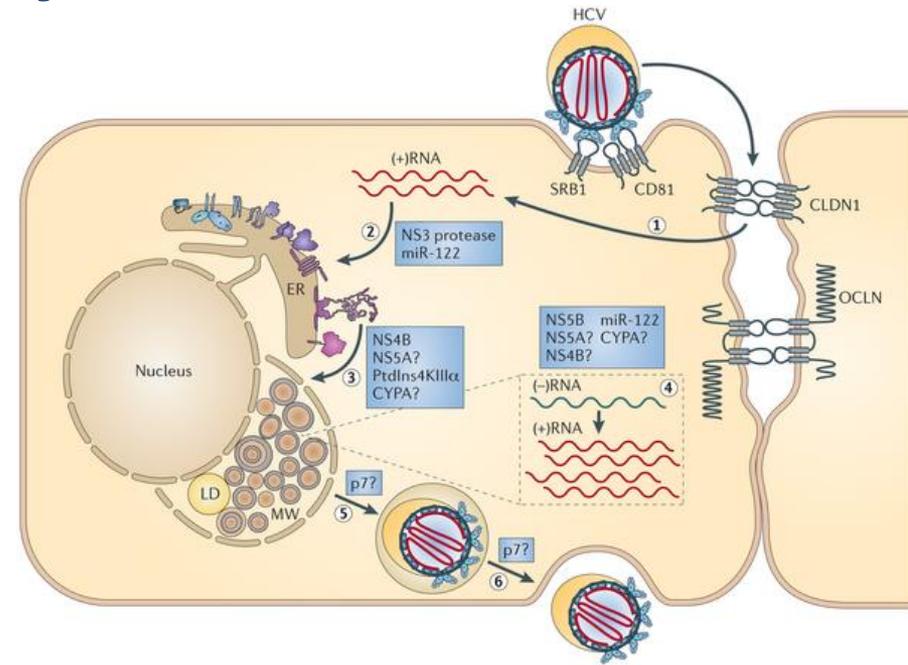
VH sequencing of sub-libraries

4. Bioinformatic analysis
5. Recovery of the whole ScFvs (VH+VL)

ELISA of the recovered clones to confirm binding

# Proof of concept: NGS on 4 cycles of selection on Claudin 1 expressing cells

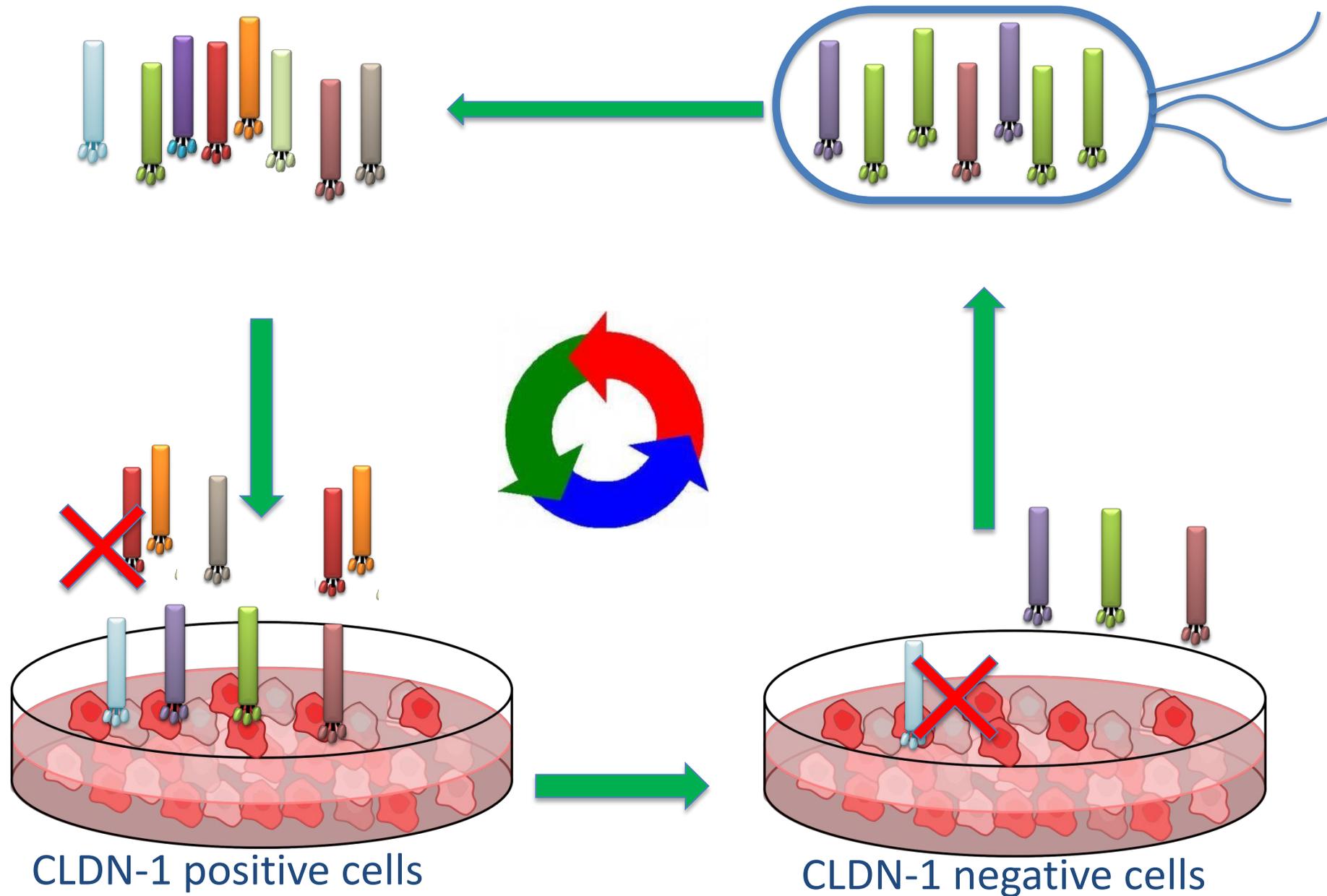
Hepatitis C virus (HCV) is a major cause of chronic hepatitis and liver carcinoma and new therapies based on novel targets are needed.



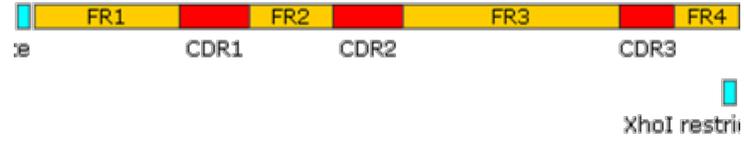
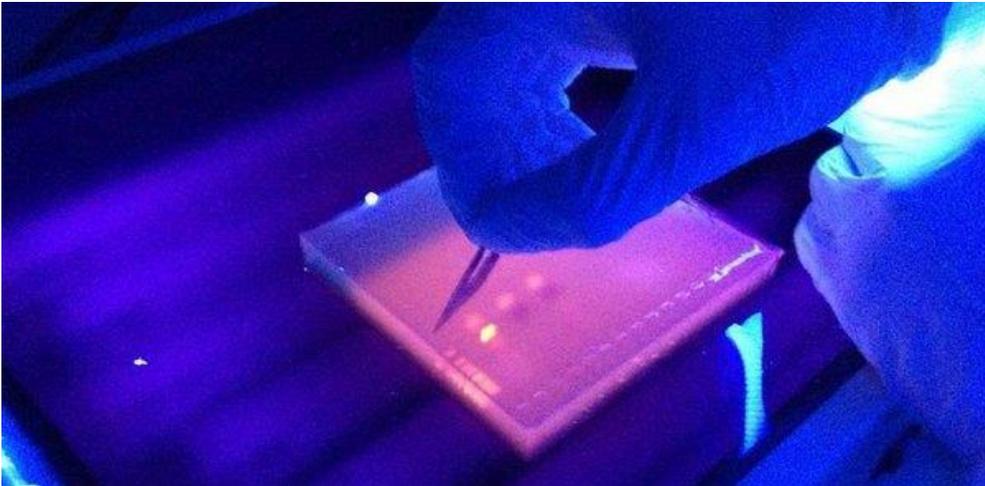
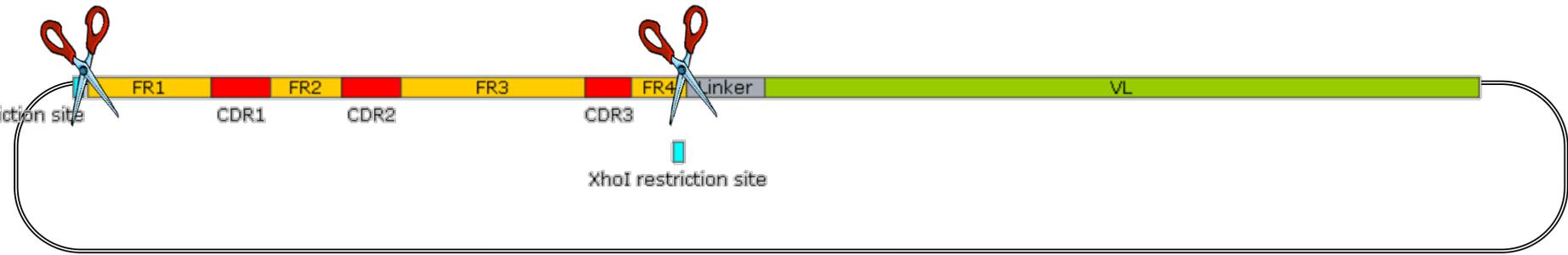
Nature Reviews | Microbiology

- The tight junction protein Claudin-1 (CLDN-1) is essential for hepatitis C virus (HCV) cell entry and spread.
- Human antibodies are not available
- We screened a phage display library of human single chain antibody fragments (scFv) by using a panel of CLDN-1-positive and –negative cell lines and identified phage specifically binding to CLDN-1.

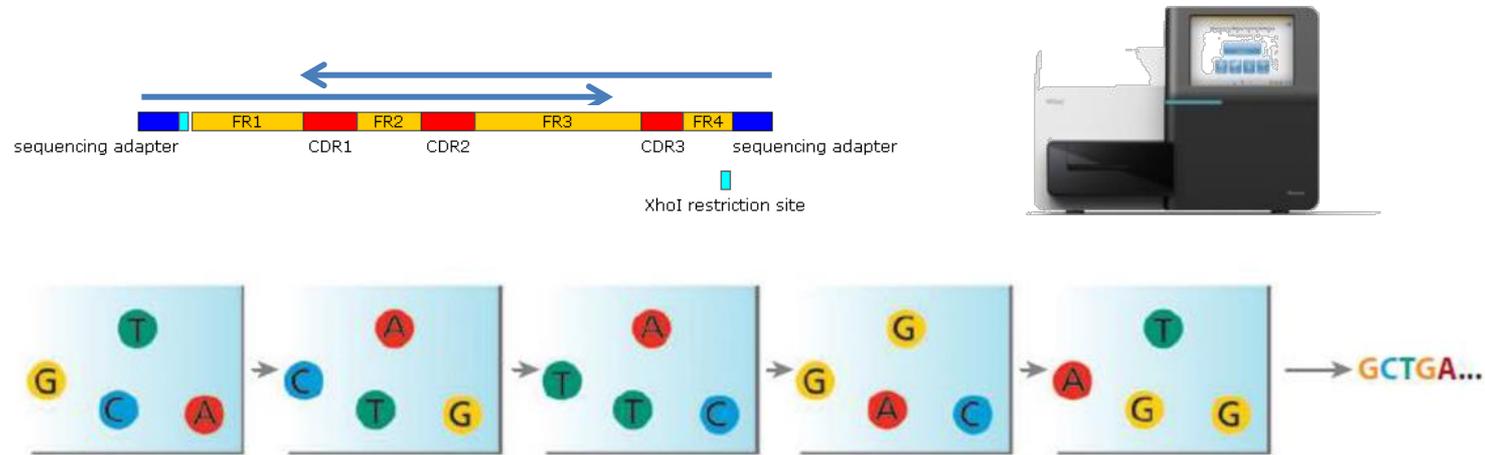
# 1. Perform 4 selection cycles by phage display



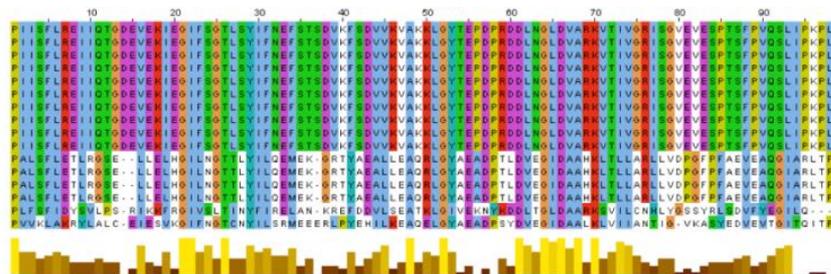
# 2. Isolation and purification of VHs from scFv plasmid



### 3. Construction of library and sequencing paired-end reads (2x300 v3)



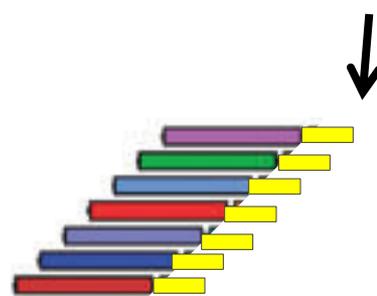
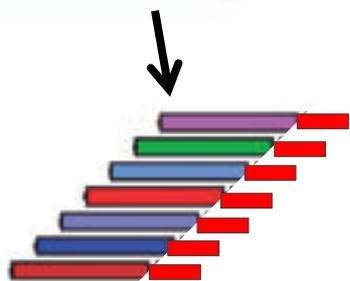
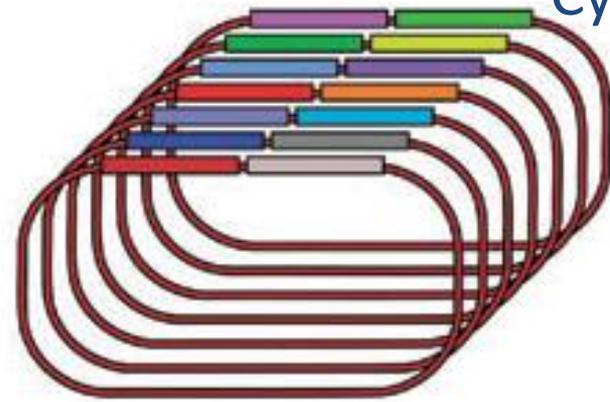
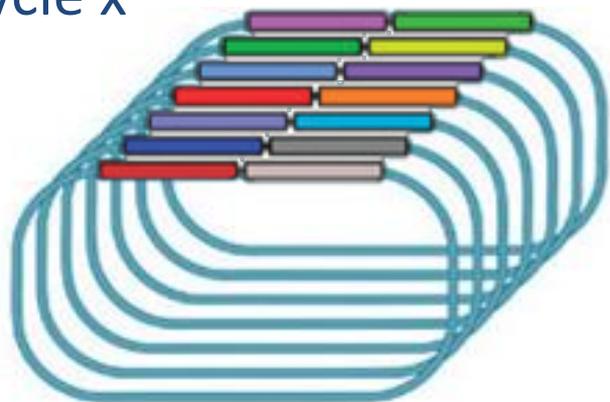
### 4. Bioinformatics: clustering sequences based on counts



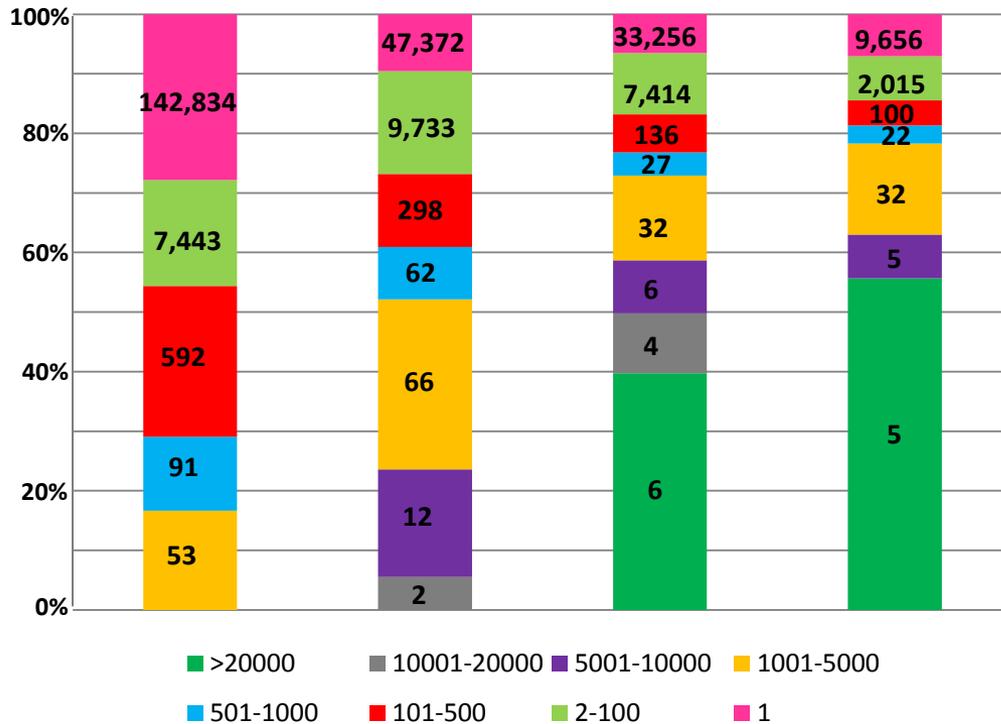
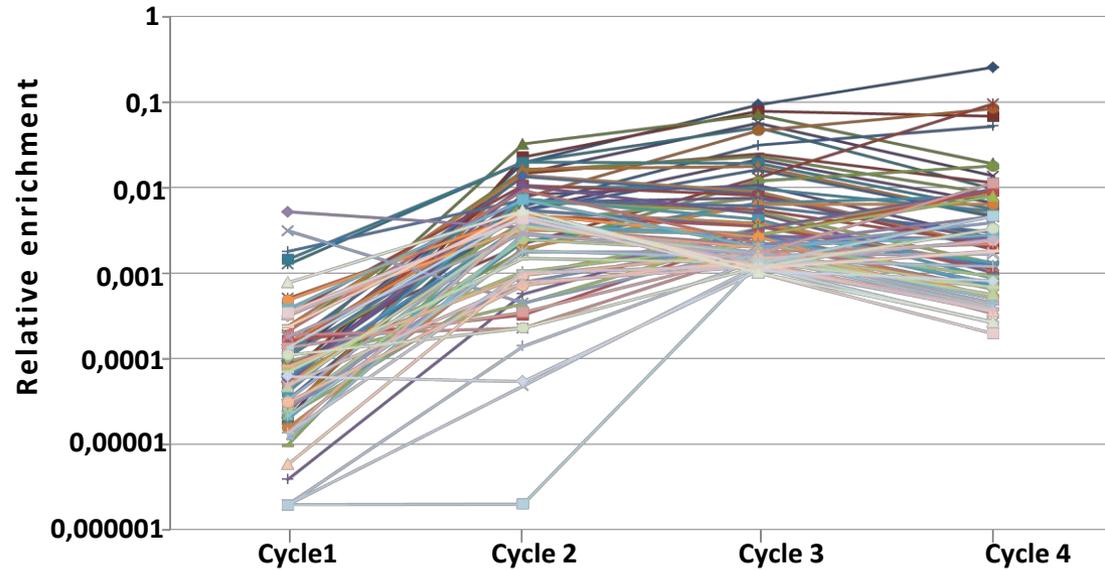
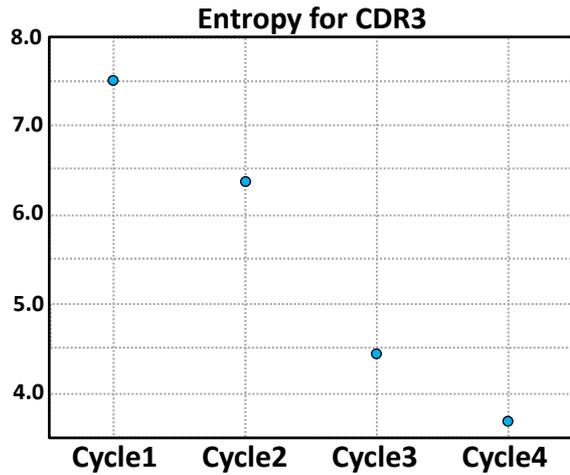
Using different barcodes it's possible to sequence VHs extracted at least from two selection cycles in the same run of sequencing

Cycle x

Cycle y



# Enrichment of sequences within and between the cycles



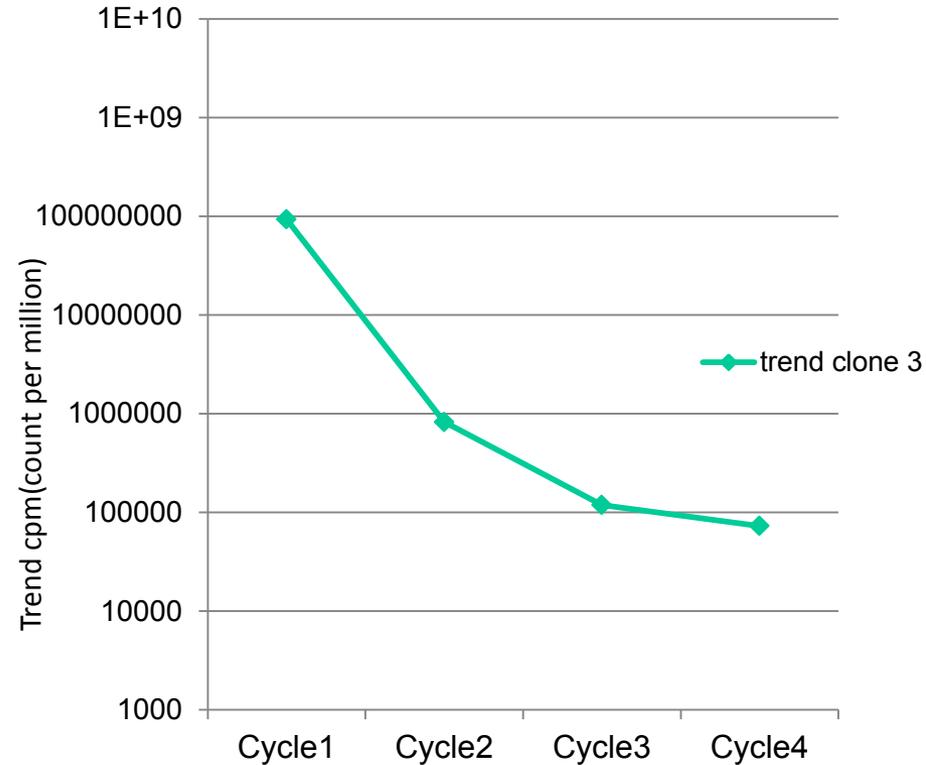
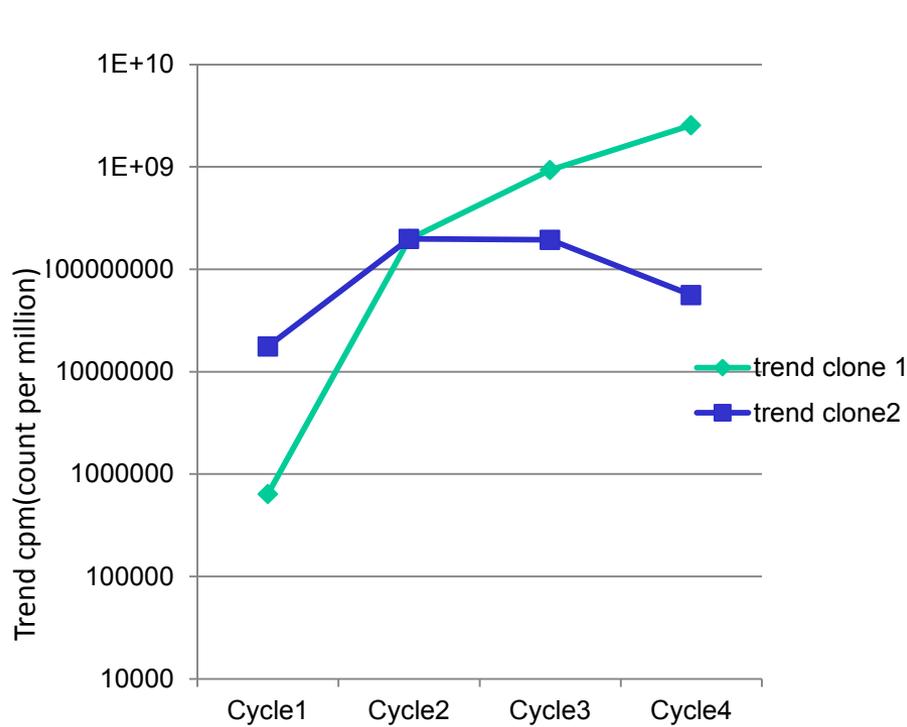
Cycle	1	2	3	4
Maximal relative enrichment	0.76%	3.24%	9,30%	25,49%
Total number of clones	151,013	57,545	40,881	11,835

# Output of bioinformatic analysis

NGS allows us to rapidly identify the potential binders, based on the counts of the corresponding VH fragment, within a cycle, and on the kinetic of their enrichments, within consecutive cycles

VH sequence	Relative abundance			
	Cycle1	Cycle2	Cycle3	Cycle4
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	638280	197490000	930730000	2549800000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1583900	21999000	131300000	960500000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	307320	76484000	465810000	847170000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	236400	229510000	790000000	684690000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	401880	61235000	314890000	523510000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1276600	324720000	714790000	190820000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	709200	18403000	119640000	180040000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1205600	147480000	570190000	137140000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1631200	3495700	16746000	113760000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	803760	146700000	247810000	113110000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	47280	5705700	33097000	96501000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	15461000	200660000	502160000	96064000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1985800	3254600	28162000	91330000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	260040	9402300	33235000	87689000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	2505800	156120000	230490000	83683000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	330960	23044000	66550000	80333000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1134700	4440000	27924000	79532000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	803760	55007000	213300000	66859000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	189120	19628000	64112000	63654000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	17612000	198210000	193660000	55934000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	1418400	102960000	97209000	53094000
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	283680	164720000	176010000	47486000
...				
GAGGTGCAGCTGTTGGAGTCTGGGGGAGGCTTGGTACAGCCTG	2788	3	0	0

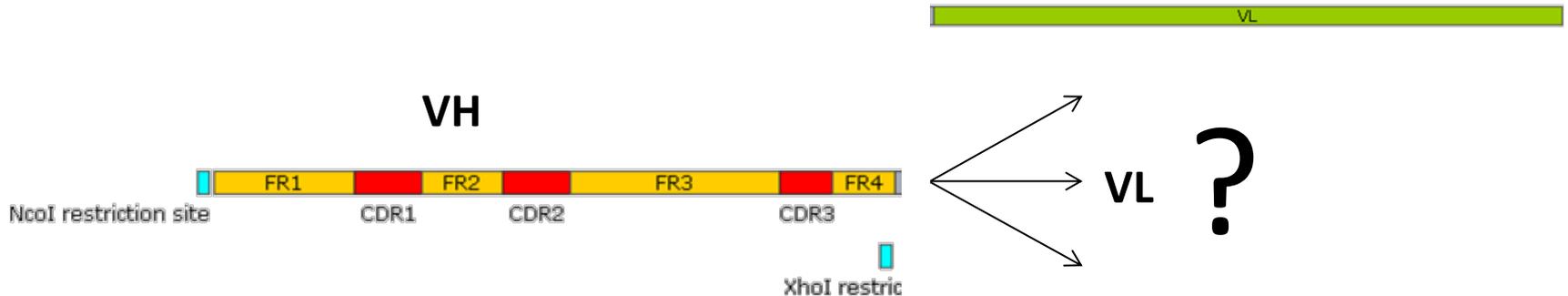
# Examples mode of enrichment or trend of several clones according to NGS analysis



- The analysis of NGS data shows that the mode of enrichment or trend could be different between several potential good binders.
- Non-specific binders could be counter-selected during selection steps.

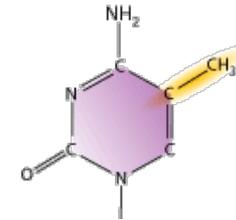
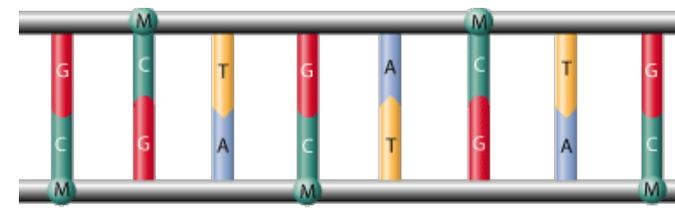
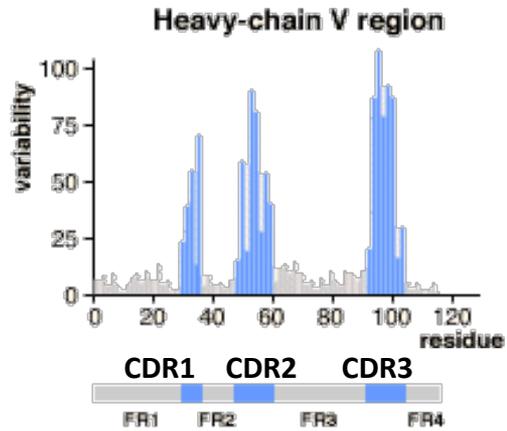
# Problems:

- Sequencing only the extracted VH, we have no information about VL sequence
- How to identify the VL associated to the enriched VH of interest?



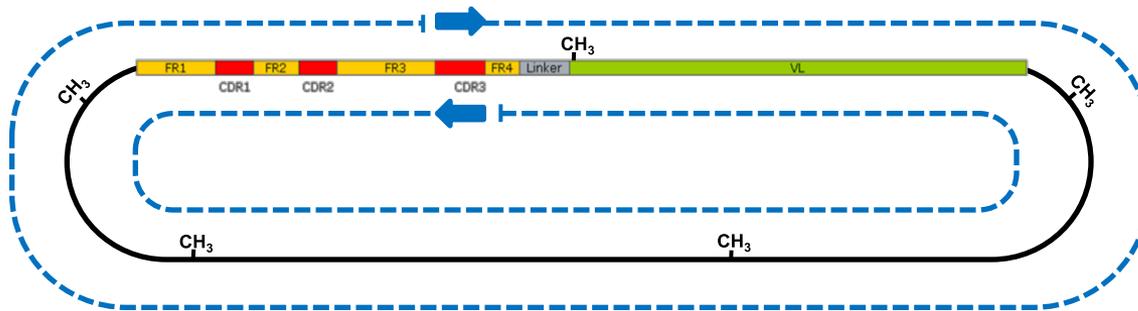


## 5. Rescue strategy of the whole ScFv (VH+VL)

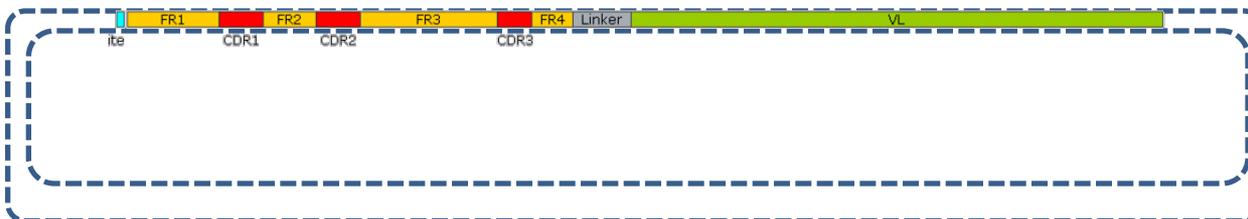


DNA methylation is the addition of a methyl group (M) to the DNA base cytosine (C).

The **CDR3** of VH is the **most variable** region of an scFv, and it's **almost unic**  
 The oligonucleotide sequences were designed inside the HCDR3 regions

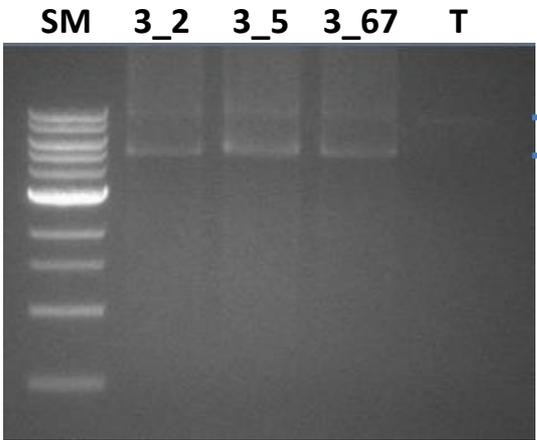


↓ Dpn1 allows to degradate the methylated and hemimethylated template DNA



# 5. Rescue strategy of the whole ScFv (VH+VL)

We decided to rescue 3 clones according to:



Growing constraint in design of specific primers

• CDR3 length:

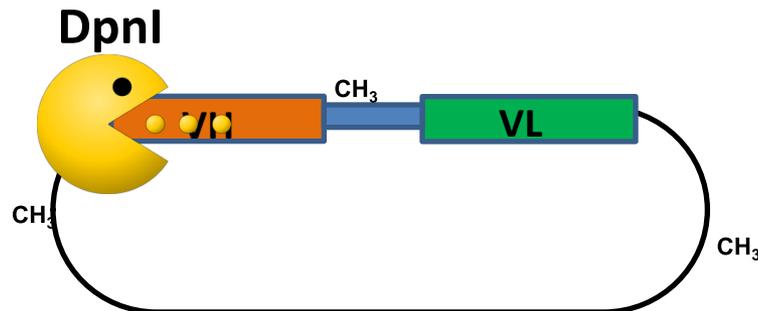
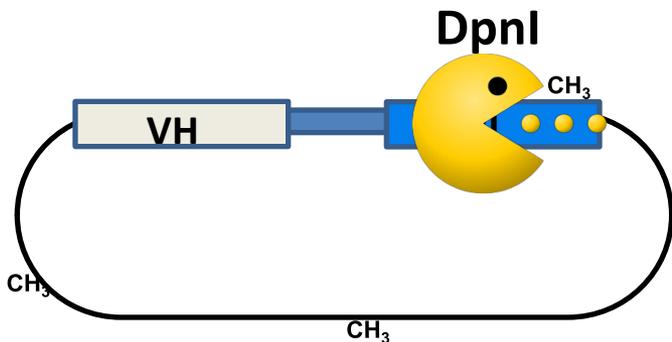
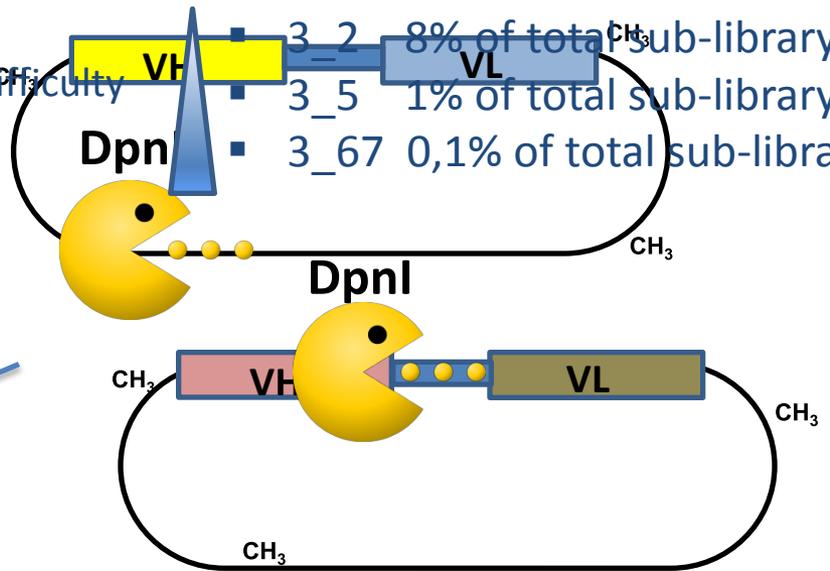
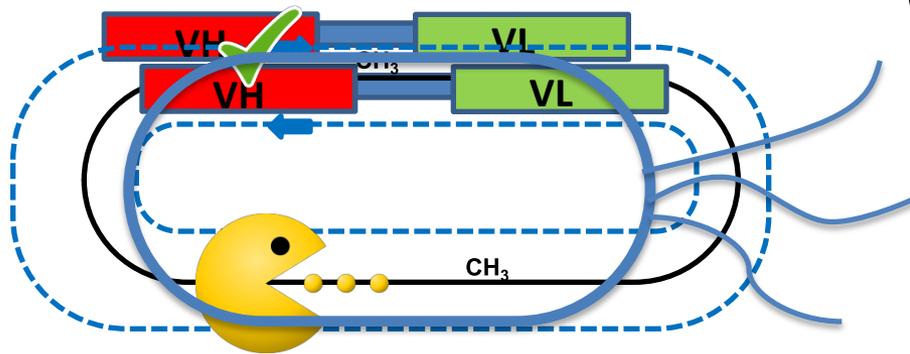
- Long CDR3 (17 a.a., clone 3\_2)
- Medium CDR3 (13 a.a., clone 3\_67)
- Short CDR3 (10 a.a., clone 3\_5)



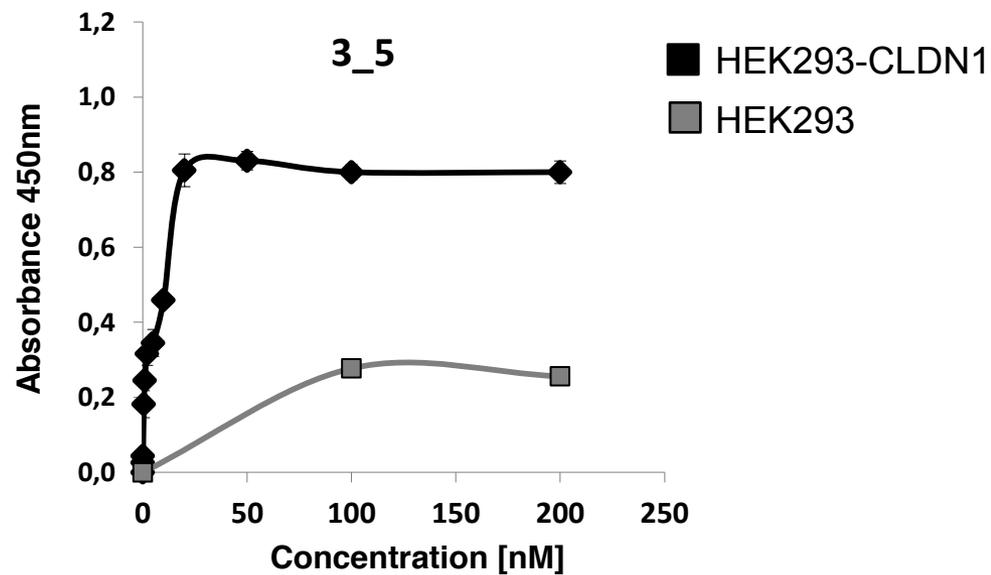
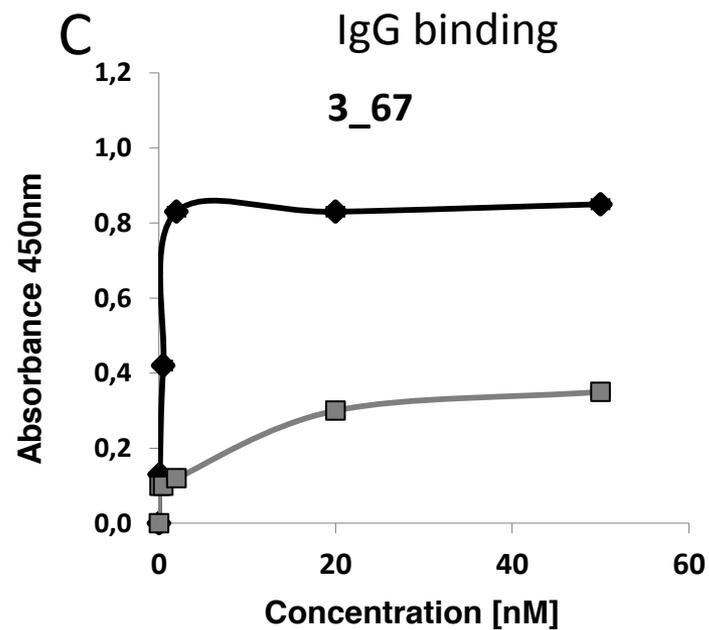
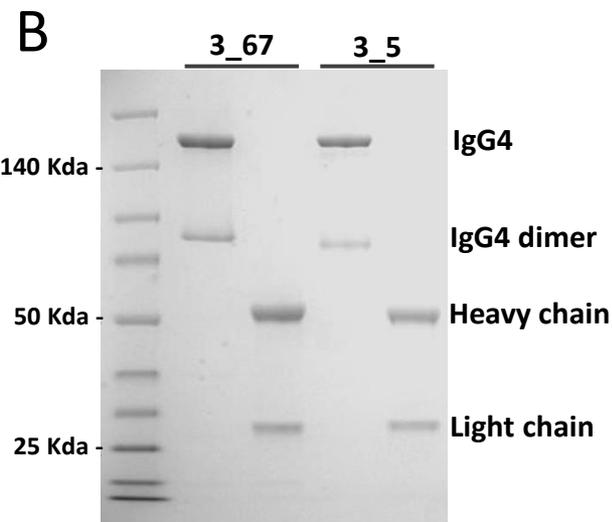
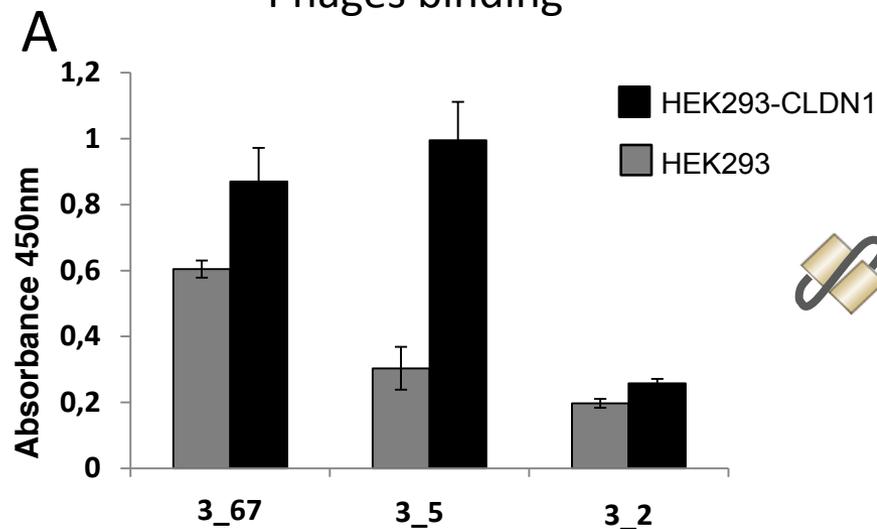
• Relative enrichment

Growing difficulty in rescue

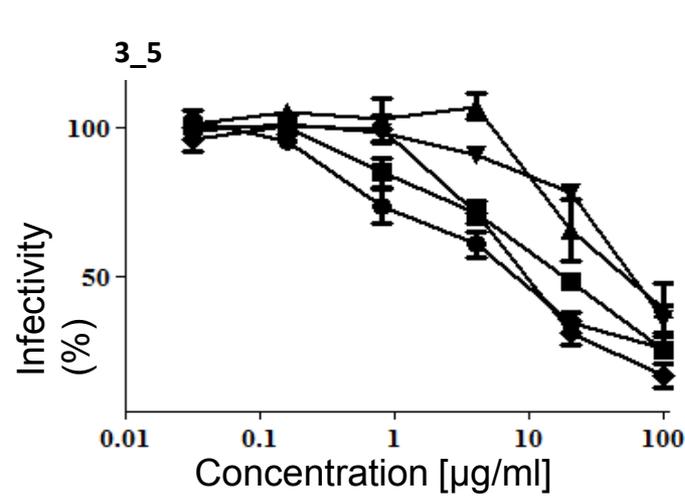
- 3\_2 8% of total sub-library
- 3\_5 1% of total sub-library
- 3\_67 0,1% of total sub-library



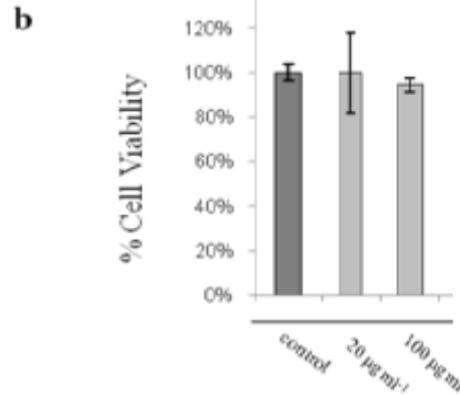
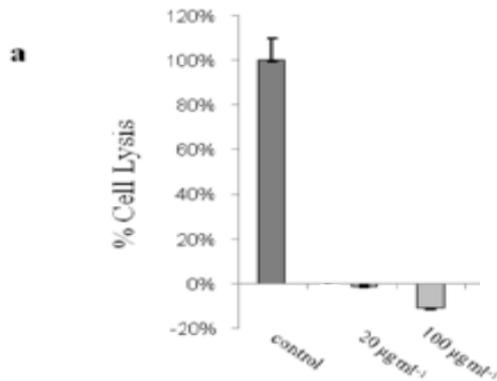
## Phages binding



# 3\_5 antibody counteracts HCV infection



Neutralization curves of 3\_5 anti-claudin 1 antibody. The mAb was tested on Huh7.5 cells treated with 2B2.8 (▼), JFH-1 (●), J6 (■), 2B1.1 (▲) and H77/JFH-1 (◆). 3\_5 has capabilities against the isolates tested.



- a) percentage of cell lysis after incubation with 3\_5 anti-CLDN-1 mAb; as positive control, cells were treated with 1% triton X-100.
- b) percentage of cell viability after 72 h incubation with 3\_5 anti-CLDN-1 mAb measured by MTT.

# Comparing the classical ELISA with NGS to identify positive scFvs during phage library selections



Antibody  
Screening

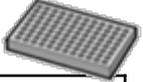
## High throughput

Enrichment evaluation and sequence analysis are contextual

Minimal handling of clones during selection steps

Sequence analysis limited to VH

Post-analysis recovery of clones



Antibody  
Screening

## Classical (ELISA)

Information in phage clones enrichment needs sequencing after each step

Repetitive isolation of the same clones within and between selection steps

Continuous handling of clones during selection steps (isolation, growth, ELISA)

Possibility to sequence the whole scFv

Positive clones recovered after each cycle

High throughput	Classical (ELISA)
Enrichment evaluation and sequence analysis are contextual	Information in phage clones enrichment needs sequencing after each step
Minimal handling of clones during selection steps	Repetitive isolation of the same clones within and between selection steps
Sequence analysis limited to VH	Continuous handling of clones during selection steps (isolation, growth, ELISA)
Post-analysis recovery of clones	Possibility to sequence the whole scFv
	Positive clones recovered after each cycle

# Conclusions

- This novel approach can guarantee rapid and cheap isolation of antibodies for virtually any antigen involved in human diseases, for therapeutic and/or diagnostic applications.
- NGS analysis of phage display libraries allows to identify good binder clones
- New potential binders are identified by NGS compared to the classical method
- Novel human anti-CLDN-1 mAbs represent useful reagents for anti-HCV therapy.

# Future perspectives

- Application of high throughput screening for isolation of new monoclonal antibodies.
- Overcome problems related to short reads of NGS technologies to obtain the whole scfv sequence.



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Thanks for attention

# Epitope or «in vivo» display?

## Phage Display Screening

